INHIBITORS OF THE MAP KINASE PATHWAY

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Field of the Invention

This invention relates to the development and use of human therapeutics that inhibit intracellular signaling via the MAP kinase pathways.

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Background of the Invention

The evolutionarily conserved Ras-MAPK signaling network regulates diverse biological processes such as cell proliferation, differentiation, migration, and survival. Many of the regulators and effectors within this network have been implicated in diverse pathological processes. MAP kinases and their targets have been identified as, for example, potent oncogenes or tumor suppressor genes and proinflammatory mediators.

Normally, the MAPK network is activated when growth factors or hormones bind to cell surface receptors. The extracellular signal is amplified and converted into an appropriate biological response. However, dysfunction of any component of the signaling pathway may result in a pathological condition.

Cancer, for example, is a disease marked by the uncontrolled growth of abnormal cells. Cancerous cells have overcome the barriers imposed in normal cells, which have a finite lifespan, and grow indefinitely. As the growth of cancer cells continues, genetic alterations can accrue and persist so that the cancerous cell displays increasingly aggressive growth phenotypes. If left untreated, metastasis, the spread of cancer cells to distant areas of the body by way of the lymph system or bloodstream, may ensue, destroying healthy tissue and, ultimately, leading to death.

According to a recent American Cancer Society study, at least 1,268,000 new cancer cases are expected to be diagnosed in the United States in any given year.

However in cancer cells, mutations in upstream activators of MAPK, such as Ras or Raf, lead to constitutive signaling even in the absence of growth factors. Constitutively activating mutations in Ras are detected in at least 30% of all human malignancies but are present in especially high levels in colon (50%) and pancreatic cancers (90%). The activation kinetics of the ERK1/2-MAPK signaling pathway have also been associated with distinct biological outcomes. In fibroblasts, sustained ERK1/2 activation over several hours induces entry into S phase of the cell cycle while transient (20-30 min) activation does not.

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In various cell types, the ERK1/2 pathway also has a critical role in regulation of nucleotide biosynthesis, transcription, migration, cell survival, differentiation and adaptive responses. Specifically, ERK signaling can control cardiomyocyte cell growth and the response to ventricular heart failure, cell survival in atherosclerosis, various metabolic processes such as glucose uptake, protein synthesis and leptin signaling, regulation of the immune response such as in T cell activation and inflammatory cytokine signaling, and mediating the effect of neurotransmitters that control memory and behavior. ERK signaling also can control the induction of genes that are required for establishing circadium rhythms.

Accordingly, small molecule drugs that can selectively inhibit regulatory proteins within the ERK1/2-MAPK pathway have enormous therapeutic potential. General MAPK inhibitors, however, are likely to be toxic due to the many metabolic and proliferative functions regulated by this pathway in healthy cells. ERK1/2 specifically recognizes some physiological substrates through the presence of ERK1/2 docking sites in substrates (Jacobs *et al.*, 1999; Tanoue *et al.*, 2000). At least two classes of docking site have been identified and are known as the D-box and DEF domain.

Substrate docking directs ERK1/2 to phosphorylate specific amino acids known to regulate the biological function of the substrate. Interaction of ERK1/2 with the D-box docking site is required for ERK's initial activation by MEK, as well as its inactivation by phosphatases (Tanoue *et al.*, 2000). By contrast, the DEF domain appears to be mainly found in downstream targets of ERK1/2 (Jacobs et al., 1999).

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Summary of the Invention

We have discovered that MAP kinases (e.g., extracellular signal-regulated kinase 1/2 (ERK1/2)), bind to certain target proteins (e.g., immediate early gene (IEG) products) through a DEF domain. This specific binding interaction results in the phosphorylation of target residues and a resulting biological effect (e.g., progression through the cell cycle). Blocking the binding events we have identified allows treatment of a variety of human diseases where the interaction of MAP kinases with the DEF domain of the target proteins has a causative biological effect.

Accordingly, in one aspect, the present invention provides for a method of identifying therapeutic compounds that affect the MAP kinase-DEF domain interaction. The method consists of the steps of: (i) providing test cells that express a target protein having a DEF domain and a MAP kinase, and are capable of progressing through the cell cycle; (ii) culturing the test cells in the presence of a growth factor, cytokine, tumor promoter, or oncogene under conditions that activate the MAP kinase; (iii) contacting the test cells with the candidate compound; (iv) assessing the binding of said MAP kinase to the DEF domain of the target protein relative to the binding in the absence of said candidate compound, wherein a candidate compound that inhibits the binding is identified as a therapeutic compound. Desirably, the test cells are mammalian; most desirably

human. Suitable test cells include, for example, a primary cell line, an immortalized cell line, or a tumor cell line. Fibroblasts (e.g., NIH 3T3 cells) are particularly useful test cells, but any mammalian cell type can be used because IEGs are ubiquitously expressed. Useful growth factors, cytokines, tumor promoters, and oncogenes include, for example, epidermal growth factor (EGF) and EGF-related factors including, for example, transforming growth factor a (TGFα), heparin-binding-like EGF, heregulin, amphiregulin, epiregulin, cripto. platelet derived growth factor (PDGF), including PGDF-AA, PGDF-BB, and PGDF -CC, insulin, insulin-like growth factors (IGFs), fibroblast growth factors (FGFs), colony stimulating factor (CSF), and heaptocyte growth factor (HGF). Useful cytokines include, for example, the chemokines, interleukins, and lysophosphatidic acid (LPA). Useful tumor promoters include, for example, phorbol esters, phosphatase inhibitors such as okadaic acid, microcystin, vanadate, hydrogen peroxide, and calyculin A. Useful oncogenes include, for example, Erb2/neu, sis, kit, Ras, Raf, PI3-kinase, and PTEN. Epidermal growth factor (EGF) and platelet-derived growth factor (PDGF) are particularly useful growth factors. In other embodiments in which the target protein is c-Fos, the binding of the MAP kinase to c-Fos is assessed by measuring the phosphorylation of T325 or T331. Preferably, this is performed using a phospho-T325-specific antibody.

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In another aspect, the invention provides a method for identifying a therapeutic compound by (i) providing a sample that contains a polypeptide having a DEF domain, a MAP kinase, and a candidate compound, (ii) contacting the target protein, the MAP kinase, and the candidate compound, and (iii) assessing the binding of the MAP kinase to the DEF domain of the target protein in the sample in the presence of the candidate compound relative to binding in the absence of the candidate compound, wherein a compound that inhibits binding of the MAP kinase to the target protein is identified as a therapeutic compound. In desirable

embodiments, the target protein further contains a fluorescent moiety (e.g., fluorescein).

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In preferred embodiments of the previous two aspects, the MAP kinase is ERK1/2. In other desirable embodiments, the target proteins are members of the Fos, Jun, and Myc family proteins. Specifically, desirable target proteins include c-Fos, Fra-1, Fra-2, cMyc, N-Myc, JunD, JunB, c-Jun, in addition to Egr-1 and mPer1. In one embodiment, the target protein contains the sequence of a protein identified in Table 1 or 2 and the identified therapeutic is useful for treating cancer. In another embodiment, the target protein contains the sequence of a protein identified in Table 3 and the identified therapeutic is useful for treating a cardiovascular disease. In another embodiment, the target protein contains the sequence of a protein identified in Table 4 and the identified therapeutic is useful for treating an inflammatory disorder. In another embodiment, the target protein contains the sequence of a protein identified in Table 5 and the identified therapeutic is useful for treating a metabolic disorder. In another embodiment, the target protein contains the sequence of a protein identified in Table 6 and the identified therapeutic is useful for treating a neuropathy or a behavioral disorder. In another embodiment, the target protein contains the sequence of a protein identified in Table 7 and the identified therapeutic is useful for treating a sleep disorder. In other embodiments, the target protein contains a DEF domain having the amino acid sequence F/Y-X-F/Y-X (SEQ ID NO: 28), preferably the sequence is F/Y-X-F/Y-P (SEQ ID NO: 29), more preferably the sequence is F-X-F-P (SEQ ID NO: 1). Assessment of target residue phosphorylation is desirably performed using a phospho-specific antibody.

In another aspect, the invention provides a method for treating cancer in a mammal (e.g., human) by administering a therapeutically effective amount of a compound that inhibits the binding of a MAP kinase to the DEF domain of a target

protein. Preferably, the MAP kinase is ERK 1/2, and the target protein is a member of the Fos, Jun, and Myc family proteins including, for example, c-Fos, Fra-1, Fra-2, cMyc, N-Myc, JunD, JunB, and c-Jun. Alternatively, the target protein is one identified in Tables 1 or 2. Alternatively, the compound inhibits the binding of a MAP kinase to a DEF domain having the amino acid sequence F/Y-X-F/Y-X (SEQ ID NO: 28), preferably the sequence is F/Y-X-F/Y-P (SEQ ID NO: 29), more preferably the sequence is F-X-F-P (SEQ ID NO: 1).

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In another aspect, the invention provides a method for treating a cardiovascular disease in a mammal (e.g., human) by administering a therapeutically effective amount of a compound that inhibits the binding of a MAP kinase to the DEF domain of a target protein. Preferably, the MAP kinase is ERK 1/2, and the target protein is one identified in Table 3. Alternatively, the compound inhibits the binding of a MAP kinase to a DEF domain having the amino acid sequence F/Y-X-F/Y-X (SEQ ID NO: 28), preferably the sequence is F/Y-X-F/Y-P (SEQ ID NO: 29), more preferably the sequence is F-X-F-P (SEQ ID NO: 1).

In another aspect, the invention provides a method for treating an inflammatory disorder in a mammal (e.g., human) by administering a therapeutically effective amount of a compound that inhibits the binding of a MAP kinase to the DEF domain of a target protein. Preferably, the MAP kinase is ERK 1/2, and the target protein is one identified in Table 4. Alternatively, the compound inhibits the binding of a MAP kinase to a DEF domain having the amino acid sequence F/Y-X-F/Y-X (SEQ ID NO: 28), preferably the sequence is F/Y-X-F/Y-P (SEQ ID NO: 29), more preferably the sequence is F-X-F-P (SEQ ID NO: 1).

In another aspect, the invention provides a method for treating a metabolic disorder in a mammal (e.g., human) by administering a therapeutically effective amount of a compound that inhibits the binding of a MAP kinase to the DEF domain of a target protein. Preferably, the MAP kinase is ERK 1/2, and the target protein is one identified in Table 5. Alternatively, the compound inhibits the binding of a MAP kinase to a DEF domain having the amino acid sequence F/Y-X-F/Y-X (SEQ ID NO: 28), preferably the sequence is F/Y-X-F/Y-P (SEQ ID NO: 29), more preferably the sequence is F-X-F-P (SEQ ID NO: 1).

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In another aspect, the invention provides a method for treating a neuropathy or a behavioral disorder in a mammal (e.g., human) by administering a therapeutically effective amount of a compound that inhibits the binding of a MAP kinase to the DEF domain of a target protein. Preferably, the MAP kinase is ERK 1/2, and the target protein is one identified in Table 6. Alternatively, the compound inhibits the binding of a MAP kinase to a DEF domain having the amino acid sequence F/Y-X-F/Y-X (SEQ ID NO: 28), preferably the sequence is F/Y-X-F/Y-P (SEQ ID NO: 29), more preferably the sequence is F-X-F-P (SEQ ID NO: 1).

In another aspect, the invention provides a method for treating a sleep disorder in a mammal (e.g., human) by administering a therapeutically effective amount of a compound that inhibits the binding of a MAP kinase to the DEF domain of a target protein. Preferably, the MAP kinase is ERK 1/2, and the target protein is one identified in Table 7. Alternatively, the compound inhibits the binding of a MAP kinase to a DEF domain having the amino acid sequence F/Y-X-F/Y-X (SEQ ID NO: 28), preferably the sequence is F/Y-X-F/Y-P (SEQ ID NO:

29), more preferably the sequence is F-X-F-P (SEQ ID NO: 1). 25

Particularly useful DEF domain inhibitors for any of the therapeutic methods are polypeptides having the sequence F/Y—X—F/Y—X (SEQ ID NO: 28; "naked DEF domains") and chimeric proteins that contain a DEF domain inserted into a non-target protein. In preferred embodiments, the DEF domain has the sequence F/Y-X-F/Y-P (SEQ ID NO: 29), more preferably the sequence is F-X-F-P (SEQ ID NO: 1). The most desirable chimeric proteins are based on non-target proteins that affect the pharmacokinetic or pharmacodynamic properties compared to administering the naked DEF domain alone. For example, DEF domains may be incorporated into serum albumin or cereloplasmin.

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The compound is administered in an amount, frequency, and duration that is therapeutically effective for treating the diagnosed condition. Desirably, the compound is administered in an amount between 0.01 and 3000 mg/day, more preferably, in an amount between 0.1 and 2000 mg/day, either orally or by injection (i.e., intravenous, intramuscular, or subcutaneous). Alternatively, the compound can be administered as a 0.5% to 25% topical formulation.

Therapy may be provided in any appropriate location: at home, the doctor's office, a clinic, a hospital's outpatient department, or a hospital. Treatment generally begins at a hospital so that the doctor can observe the therapy's effects closely and make any adjustments that are needed. The duration of the therapy depends on the condition being treated, the age and condition of the patient, the stage and type of the patient's disease, and how the patient's body responds to the treatment. Drug administration may be performed at different intervals (e.g., daily, weekly, or monthly).

In another aspect, the invention provides an antibody that specifically binds to phospho-T-325 c-Fos. The antibody may be monoclonal or polyclonal.

In another aspect, the invention provides a pharmaceutical formulation that contains a therapeutic compound identified by either of the first two aspects of the invention, and a pharmaceutically acceptable carrier. The pharmaceutical formulation may be suitable for oral administration, injection, or topical application.

By "specifically binds to phospho-T-325 c-Fos," when referring to the antibodies of this invention, is meant an antibody that binds with high affinity (<10⁻⁸M) to native c-Fos in which the threonine at amino acid position 325 is phosphorylated, but does not significantly bind to c-Fos in which the T325 is unphosphorylated. Desirably, the difference in specificity of antibody binding between phospho-T-325 c-Fos and the unphosphorylated form is at least 5-fold, 10-fold, 25-fold, 50-fold, 100-fold, or 1000-fold.

By "DEF domain" is meant a polypeptide having the amino acid sequence: $F/Y-X_1-F/Y-X_2$ (SEQ ID NO: 28), wherein F is phenylalanine, Y is tyrosine, P is proline, and X_1 and X_2 are any naturally-occurring or non-naturally-occurring amino acids. Desirably, X_2 is proline.

By "target protein" is meant any protein that contains a DEF domain capable of binding a target kinase (e.g., a MAP kinase). Desirable target proteins are phosphorylated by the MAP kinase ERK1/2 following ERK1/2 binding to the DEF domain. Target proteins include, for example, gene products of the immediate early genes from the Fos, Myc, and Jun families, proteins identified in Tables 1-7, or chimeric or synthetic proteins into which a DEF domain has bee inserted by artifice. Specific target proteins include, for example, c-Fos, Fra-1, Fra-2, cMyc, N-Myc, JunD, JunB, c-Jun, Egr-1, and mPer1.

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By "target residue(s)" is meant one or more residues of a target protein that are N-terminal to the DEF domain and that are phosphorylated as a result of the binding of a MAP kinase. Target residues include, for example, T325 and T331 of c-Fos. This phosphorylation event is also termed a "DEF domain-dependent phosphorylation."

By "primed," when referring to a target protein, is meant a phosphorylation event that makes a DEF domain available for binding of a MAP kinase. Thus, the amino acid residues that are the subject of a "priming" modification are not the same as the target residues. For example, c-Fos is primed when S362 and/or S374 are phosphorylated or substituted for aspartate or glutamate.

By "target kinase" is meant a protein kinase that is capable of binding a DEF domain and phosphorylating a target residue. Target kinases include the MAP kinases such as ERK1/2, for example. Thus, an "activated target kinase" is one that itself has undergone a post-translational modification causing an increase in kinase activity and/or inducing a change in subcellular localization. For example, in order to be fully activated and translocated from the cytoplasm to the nucleus, ERK1/2 is phosphorylated.

By "DEF domain inhibitor" is meant any chemical compound (i.e., polypeptide or non-peptide) that inhibits the interaction of a target kinase (i.e., ERK1/2 or RSK) with the DEF domain of a target protein.

The term "assessing the binding of a MAP kinase to a DEF domain," is meant to include any appropriate binding or biochemical assessment which may be either qualitative or quantitative. This term specifically includes, for example, directly assessing the interaction of the MAP kinase and the DEF domain.

Alternatively, assays that measure biochemical outcomes of a MAP kinase – DEF domain binding event are useful. These assays include, for example, measuring the amount of DEF domain-dependent phosphorylation.

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As exemplified in detail below, the phosphorylation of T325 and/or T331 of c-fos is dependent upon this binding event.

By "cancer" is meant neoplastic cells multiplying in an abnormal manner. In a cancer, growth is uncontrolled and progressive, and occurs under conditions that would not elicit, or would halt the multiplication of non-cancerous cells. Cancer includes, for example, leukemias and lymphomas (Hodgkin's disease, non-Hodgkin's disease), as well as solid tumors such as sarcomas and carcinomas (e.g., fibrosarcoma, liposarcoma, osteogenic sarcoma, colon carcinoma, pancreatic cancer, breast cancer, ovarian cancer, prostate cancer, squamous cell carcinoma, basal cell carcinoma, adenocarcinoma, renal cell carcinoma, hepatoma, bile duct carcinoma, Wilm's tumor, cervical cancer, uterine cancer, testicular cancer, small and/or non-small cell lung carcinoma, bladder carcinoma, epithelial carcinoma, glioma, astrocytoma, medulloblastoma, schwannoma, meningioma, melanoma, neuroblastoma, and retinoblastoma.

By "treating cancer" is meant a therapy that measurably slows, stops, or reverses the growth rate of the cancer (i.e., neoplastic cells) in vivo. Desirably, a slowing of the growth rate is by at least 20%, 30%, 50%, or even 70%, as determined using a suitable assay for determination of cell growth rates (e.g., a cell growth assay described herein). Typically, a reversal of growth rate is accomplished by initiating or accelerating necrotic or apoptotic mechanisms of cell death in the neoplastic cells, resulting in a shrinkage of the neoplasm. Efficacy of a treatment may be measured by any means known to those skilled in the art including tumor imaging or measurement of neoplastic markers.

By "cardiovascular disease" is meant ischemic heart disease, ventricular heart failure, cardiac hypertrophy, hypertension, and atherosclerosis.

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By "inflammatory disorder" is meant any condition that is characterized by inflammation as a primary or secondary symptom. Inflammatory disorders include, for example, allergic or autoimmune disorders, anaphylaxis, and septic shock. Examples of allergic disorders include allergic rhinitis, asthma, atopic dermatitis, and food allergies. Examples of autoimmune disorders include, but are not limited to, type 1 insulin-dependent diabetes mellitus, inflammatory bowel disease, Crohn's disease, ulcerative colitis, dermatitis, meningitis, thrombotic thrombocytopenic purpura, Sjögren's syndrome, encephalitis, uveitis, leukocyte adhesion deficiency, rheumatoid and other forms of immune arthritis, rheumatic fever, Reiter's syndrome, psoriatic arthritis, progressive systemic sclerosis, primary biliary cirrhosis, pemphigus, pemphigoid, necrótizing vasculitis, myasthenia gravis, multiple sclerosis, lupus erythematosus, polymyositis, sarcoidosis, granulomatosis, vasculitis, pernicious anemia, CNS inflammatory disorder. antigen-antibody complex mediated diseases, autoimmune hemolytic anemia. Hashimoto's thyroiditis, Graves disease, habitual spontaneous abortions, Reynard's syndrome, glomerulonephritis, dermatomyositis, chronic active hepatitis, celiac disease, autoimmune complications of AIDS, atrophic gastritis, ankylosing spondylitis and Addison's disease.

By "metabolic disorder" is meant a disease that interferes with the normal metabolic function of cells, tissues or organs. Metabolic disorders include, for example, diabetes, obesity, jaundice, polycystic kidney and hepatic disease, pancreatitis, Graves' disease, and Werner's syndrome. Metabolic diseases may also arise as secondary complications of another disease such as one involving a tumor. For example, cachexia or muscle wasting, and metabolic and digestive complications often arise from the presence of pancreatic, colonic, stomach, hepatic and hepatocellular tumors.

By "neuropathy" is meant any condition of the central or peripheral nervous system characterized by axonal loss that may or may not be accompanied by neuronal loss. Neuropathies specifically include conditions affecting sensory and motor neurons and include, for example, diabetic neuropathy, muscular dystrophy, Williams Beuren's Syndrome.

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By "behavioral disorder" is meant any condition affecting motivation, emotion, learning, or memory. Behavioral disorders are also meant to broadly encompass neurodegenerative diseases. Thus, behavioral disorders include, for example, psychosis, schizophrenia, autism, Down's Syndrome, Parkinson's Disease, Alzheimer's Disease, epilepsy, Cockayne syndrome, bipolar disorders, and depression. Also included are addictions including, for example, addictions to opiates and barbiturates.

By "sleep disorder" is meant any condition that primarily affects sleep and consciousness. Sleep disorders include, for example, advanced sleep phase syndrome, delayed sleep phase syndrome, insomnia and narcolepsy.

By "a therapeutically effective amount" is meant the amount of a compound required to treat cancer (i.e., inhibit the growth of the neoplastic cells). The effective amount of active compound(s) used to practice the present invention for therapeutic treatment of neoplasms (i.e., cancer) varies depending upon the manner of administration, the age, body weight, and general health of the subject. Ultimately, the attending physician or veterinarian will decide the appropriate amount and dosage regimen.

Brief Description of Drawings

FIGURE 1 is a series of photomicrographs showing the differential responsiveness of Swiss 3T3 fibroblasts to growth factors. FIGURE 1A shows quiescent Swiss 3T3 cells (-) that were treated with EGF (25 ng/ml) or PDGF (20

ng/ml) for 20 h and then processed for BrdU incorporation, as described below. FIGURE 1B shows quiescent Swiss 3T3 cells that were treated with PDGF or EGF for the indicated times and ERK1/2 and RSK kinase activities were determined using immunecomplex kinase assays. The fold activation at each time is indicated above each lane. FIGURE 1C is the indirect immunofluorescence detection of hyperphosphorylated activated ERK1/2 in Swiss 3T3 cells treated with EGF or PDGF. FIGURE 1D is the indirect immunofluorescence detection of c-Fos in Swiss 3T3 cells treated with EGF or PDGF.

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FIGURE 2A is an illustration showing the residues in c-Fos that are phosphorylated by RSK and ERK1/2 in vivo. FIGURE 2B shows the electrophoretic separation of cell extracts from parallel cultures of 208F fibroblasts stably expressing Fos-WT (WT), Fos-AA (AA) or Fos-DD (DD) that were metabolically labelled with ³⁵S-methionine or ³²P-orthophosphate and cultured with or without 10% FBS for 15 min. Fos proteins were immunoprecipitated from cell lysates and analysed by SDS-PAGE. The serum-stimulated phosphorylation of Fos-DD was consistently two to threefold greater than Fos-AA and arrows indicate the major mobilities observed after stimulation. FIGURE 2C is Western blot from NIH 3T3 cells transfected with Fos-WT (WT), Fos-AA (AA) or Fos-DD (DD) were serum-starved and then pre-treated with 5 μM UO126 (+) or 0.1% DMSO (-) for 30 min before treatment with EGF (+) for 5 min. FIGURE 2D is an autoradiogram of an in vitro phosphorylation of the indicated (His)₆-Fos proteins by endogenous ERK1/2 from quiescent or EGF-stimulated NIH 3T3 cells. Results shown are representative of three independent experiments. Fos-EE (S362E/S374E) was used as primed c-Fos for in vitro phosphorylation studies.

FIGURE 3A is an illustration that details the DEF domain at the C-terminus of c-Fos. The phosphorylation of Fos-EE in the absence of peptide competitor is expressed as 100%. *In vitro* phosphorylation of (His)₆-Fos-EE was performed as

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described below. The data shown are the means ±SEM from three experiments. FIGURE 3B is a graph showing the inhibition of Fos-EE phosphorylation by peptides containing a DEF domain (FQFP; SEQ ID NO: 3) or a mutated DEF domain (AQAP; SEQ ID NO: 4). FIGURE 3C is a graph showing the inhibition of Fos-EE phosphorylation by peptides containing the c-Fos DEF domain (FTYP; SEQ ID NO: 2) or a mutated DEF (ATYP; SEQ ID NO: 5). FIGURE 3D shows the results of a Western blot of NIH 3T3 cells transfected with the indicated FLAG-Fos-DD (DD) alleles were left quiescent (-) or were stimulated (+) with EGF for 5 min before lysis. Arrows indicate the major Fos-DD mobilities. FIGURE 3E is a Western blot of cells transfected with the indicated FLAG-Fos alleles. Arrows show the three major Fos-WT mobilities associated with growth factor stimulation.

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FIGURE 4A is an illustration identifying the ERK1/2 phosphorylation sites N-terminal to the DEF domain in c-Fos. *In vitro* phosphorylation of (His)₆-Fos-EE proteins by activated (His)₆-ERK1/2 was performed. The phosphorylation of the Fos-EE point mutants is expressed as a percentage of Fos-EE (100%). The data shown are the means ±SEM from three experiments. FIGURE 4B is a Western blot of NIH 3T3 cells that were transfected with the indicated FLAG-Fos-DD (DD) alleles. Cells were treated with EGF for 5 min or left untreated. FIGURE 4C is a Western blot of cells transfected with the indicated FLAG-Fos alleles and treated as in Figure 3B.

FIGURE 5A is an illustration identifying the phospho-Thr 325 peptide used to generate the phospho-Thr-325-specific anti-c-Fos antiserum. FIGURE 5B is a Western blot of NIH 3T3 cells that were transfected with the indicated c-Fos alleles or with vector alone. Extracts were prepared from quiescent (-) or EGF-stimulated (+) cells and analyzed using either the anti-c-Fos antibody or the phospho-Thr 325 antiserum. Results shown are representative of three

independent experiments. FIGURE 5C is a Western blot of ΔB-Raf-ER NIH 3T3 cells transfected with Fos-WT or Fos-AA that were either starved and left untreated (0) or treated with 1 μM tamoxifen (TAMX) for the indicated times before lysis. The *in vivo* phosphorylation of Thr 325 in Fos-WT and Fos-AA was analyzed by western blotting using the phospho-Thr 325-specifi antiserum. FIGURES 5D and 5E are Western blots demonstrating the *in vivo* mitogen-regulated phosphorylation of Thr 325 in the context of the Fos DEF domain mutants. NIH 3T3 cells expressing the indicated Fos proteins were treated as in the same manner as for FIGURE 3B, and extracts analyzed for phosphorylation of Thr 325.

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FIGURE 6A is a Western blot of quiescent Swiss 3T3 cells were treated with EGF (25 ng/ml) for the indicated times. Lysates were probed for endogenous c-Fos, Thr 325 phosphorylation in c-Fos. FIGURE 6B is a Western blot of Swiss 3T3 cells that were treated with PDGF-BB (20 ng/ml) and processed as in described for Figure 6A. Results shown are representative of three experiments. FIGURE 6C is a Western blot of quiescent Swiss 3T3 cells that were treated with PDGF for 60 min and then treated with UO126 (5 μM), as indicated, or with DMSO (lanes 3-8) for the remainder of the experiment. The expression and phosphorylation of endogenous c-Fos was visualized as in above. FIGURE 6D is an autoradiogram showing the kinase activities of endogenous ERK1/2 and RSK in cell lysates from Figure 6C. The fold activity is provided above each lane.

FIGURE 7A is a bar graph showing the AP-1 transcriptional activity of the indicated c-Fos alleles in Hela cells. AP-1 luciferase activity in cells expressing Fos-WT is expressed as 100%, and the data shown are from five individual experiments. FIGURE 7B is a photomicrograph showing the expression of endogenous c-Fos in quiescent (-) or serum-stimulated (+) pMV7-infected 208F fibroblasts (vector), assayed by immunofluorescence microscopy. To examine the

expression of Fos-WT, Fos^{Y45A} or Fos^{T325A/T331A} in G418-resistant 208F fibroblasts, cells were serum-starved for 24 h before fixation and processing for immunofluorescence microscopy using the anti-Fos antibody. FIGURE 7C is a Western blot of c-Fos protein expression in quiescent 208F cells. FIGURE 7D is a bar graph showing the anchorage-independent growth of G418-resistant pools of 208F cells stably expressing pMV7 (vector) or the indicated FLAG-Fos alleles. The data are expressed as a percentage of the number of colonies formed by cells expressing Fos-WT (100%) and represent the mean ±SEM from six experiments performed in duplicate.

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FIGURE 8 is a schematic diagram of the molecular interpretation of ERK1/2 signal duration. Growth factor stimulation (stimulus) causes activation of signaling pathways (signals) that result in rapid transcriptional induction of immediate early genes (response). The duration of ERK1/2 signaling is then interpreted by immediate early gene products that contain DEF domains (signal sensors). ERK1/2-docking to the DEF domain results in sensor phosphorylation. Docking and phosphorylation alters its biological activity, and this dictates the biological outcome. TF, transcription factor.

FIGURE 9A is a photomicrograph showing the nuclear accumulation of active ERK1/2 and c-Fos in growth factor-treated Swiss 3T3 cells. These photomicrographs are enlargements of the images of Figure 1C in order to visualize the cellular distribution of activated ERK1/2 and nucleolar structures. FIGURE 9B is a photomicrograph of quiescent Swiss 3T3 cells treated with PDGF for 75 minutes followed by the addition of cyclohexamide (+) or vehicle (-). Cells were processed for c-Fos immunofluorescence 90, 180 or 300 minutes after PDGF stimulation. In control experiments (bottom two panels), cells were incubated with cyclohexamide or vehicle for 20 minutes prior to treatment with PDGF for 90 minutes.

FIGURE 10A is a Western blot of NIH3T3 cells transiently transfected with FosWT or FosDD were left quiescent or treated with EGF (50 ng/ml, 5 min) prior to lysis. An aliquot from each cell extract was incubated in the presence or absence of λ protein phosphatase (P'ase) for 30 minutes on ice. Data shown is representative of three separate experiments. FIGURE 10B is a Western blot of NIH3T3 cells stably expressing ΔB -Raf:ER that were transfected with FosWT, FosAA or FosDD. Cells were deprived of serum growth factors, pre-treated with 5 μM UO126 (+) or 0.1% DMSO (-) for 30 minutes prior to treating with tamoxifen (TAMX, 1μM) for 15 minutes prior to cell lysis. FIGURE 10C is a Western blot showing the phosphorylation of (His)₆-FosWT, AA or EE or MBP by Flag-ERK5. Active (+) and inactive ERK5 (-) was obtained by coexpressing Flag-ERK5 and HA-MEK5(D) or control vector, respectively, in 293 cells followed by immunoprecipitation of Flag-ERK5 from cell lysates using the M2 anti-Flag monoclonal antibody. Autophosphorylation (auto-P) of ERK5 in kinase reactions is indicated. Together, these data demonstrate that the phosphorylation of primed c-Fos is regulated by the Raf/Mek/ERK pathway.

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FIGURE 11 is a Western blot of quiescent Rat-1 cells that were treated with the indicated concentrations of LPA for various times. The activation kinetics of ERK1/2 demonstrates that c-Fos is a sensor for sustained ERK1/2 signaling in Rat-1 fibroblasts. The data shown is representative of at least three individual experiments.

FIGURE 12 is a series of cell culture plates, fixed and then stained with Giemsa to visualize foci. The indicated Fos proteins were stably expressed in 208F cells and cultured for four weeks in regular culture medium. Identical data was obtained from five separate experiments. Thus, substituting aspartic acid for T235 and T331 in c-Fos promotes Fos-mediated transformation.

FIGURES 13A-C are a series of Western blots showing the regulation of ectopically expressed Fos family proteins (c-Fos, Fra-1, and Fra-2) by the ERK1/2 pathway in NIH 3T3 cells. Cells were treated with or without EGF (50 ng/mL) for 5 minutes prior to cell lysis. Where indicated, UO126 (5 mM) was added to cells 30 minutes before adding EGF. EGF treatment in the absence of UO126 activated ERK1/2, as demonstrated by the mobility shift to a higher molecular weight. ERK1/2 activation resulted in phosphorylation of the target amino acid, T325, of c-Fos (Figure 13A). ERK1/2 activation of Fra-1 (Figure 13B) and Fra-2 (Figure 13C) is also demonstrated by the observed mobility shift.

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FIGURE 14A is a sequence alignment of c-Fos, Fra-1, and Fra-2 (SEQ ID NO: 25-27, respectively) demonstrating a high degree of sequence identity in the C-termini. Fra-1 and Fra-2 have ERK1/2 and RSK priming phosphorylation sites in addition to DEF domains. FIGURE 14B is a Western blot of NIH 3T3 cells transfected with the indicated constructs and deprived of serum growth factors for 24 hours. These results demonstrate that mutations in the DEF domains of Fra-1 and Fra-2 inhibit the ERK1/2-mediated mobility shift (compare to Figures 13B and 13C).

FIGURE 15A is a Western blot of c-Myc immunoprecipitation from NIH 3T3 cells transfected with pcDNA3 (vector) or c-Myc and deprived of serum growth factors. EGF and UO126 were used to treat cells as described in Figure 13. FIGURE 15B is a Western blot from cells transfected with the indicated c-Myc proteins. These results characterize the DEF domain in c-Myc and show that S62 phosphorylation depends on an intact DEF domain.

FIGURES 16A-F are Western blots demonstrating the kinetics of immediate early gene expression in Swiss 3T3 cells. Cells were deprived of serum growth factors and treated with EGF (25 ng/mL) or PDGF-BB (20 ng/mL) for the indicated times.

FIGURES 17A-B are Western blots demonstrating the kinetics of Egr-1, JunB, and c-Myc expression in Swiss 3T3 cells. Cells were treated as described in Figure 13. Total levels of c-Myc (Figure 17B) were detected by immunoprecipitating c-Myc prior to Western analysis.

FIGURES 18A-E are Western blots demonstrating that sustained expression of immediate early genes requires ERK1/2 activity. Serum deprived Swiss 3T3 cells were treated with PDGF-BB for 90 minutes before adding DMSO vehicle (0.1%) or UO126 (5 μ M).

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FIGURE 19A is a Western blot of cells treated with PDGF-BB for either 90 minutes (lanes 2-9) or 120 minutes (lanes 11-13) before adding DMSO vehicle (lanes 3-5) or UO126 (lanes 7-9, 12, 13). FIGURE 19B is a Western blot of cells treated with PDGF-BB for 5 hours to induce Fra-1 before adding UO126 for a further 20 or 30 minutes. These figures demonstrate that ERK1/2 signaling is required during G1 for the stabilization of c-Myc.

FIGURE 20 is a series of Western blots from Swiss 3T3 cells treated with various concentrations of PDGF-BB before lysis. These results demonstrate that IEG products act as sensors for subtle differences in ERK1/2 signal duration.

FIGURE 21A is a bar graph of the result from an *in vitro* kinase assay demonstrating ERK 1/2 activation is sensitive to small differences in growth factor (PDGF) stimulation. FIGURE 21B is a Western blot demonstrating that the c-Fos stabilization observed in Figure 20 following stimulation with 10 ng/ml PDGF is a result of ERK 1/2-dependent phosphorylation of T325. Neither long-term c-Fos stabilization (see Figure 20) nor T325 phosphorylation is observed following 4 ng/ml PDGF stimulation.

FIGURE 22 is a series of Western blots showing Fra-1 hyperphosphorylation throughout G1 requires ERK1/2 signaling.

FIGURE 23 is representative gel and the densitometric quantification of an electrophoretic mobility shift assay (EMSA) for AP-1. Swiss 3T3 cells were treated as indicated and extracted in a hypotonic lysis buffer. The nuclear fraction was isolated and aliquots mixed with a ³²P-labelled AP-1 oligonucleotide in a standard EMSA. These results demonstrate that PDGF, but not EGF, treatment significantly increases AP-1 expression and AP-1 DNA binding.

FIGURE 24 is an immunoprecipitation of extracts from 208F cells stably expressing c-Myc or c-Myc F196A and treated with cycloheximide (14 mg/mL) for the indicated times. Following immunoprecipitation of the c-Myc proteins, total levels of c-Myc were detected using Western analysis. These results demonstrate that c-Myc stability requires the DEF domain.

FIGURE 25 is a series of indirect immunofluorescence photomicrographs demonstrating typical results of the screening assays described herein.

Representative fields of view using a 10X objective lens are shown.

FIGURE 26 is a series of Western blots and accompanying densitometric analysis showing the effect of mutating the DEF domain binding site in ERK1/2 on a RSK (non-DEF domain-dependent) phosphorylation and c-Fos (DEF domain-dependent) phosphorylation. The bar graphs represent the raw densitometric analysis, unadjusted for ERK1/2 content.

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Detailed Description

We have discovered that DEF domains are present in numerous proteins that are important in a variety of human diseases and, by blocking the interaction of a MAP kinase with the DEF domain of a target protein, effective therapy may be provided. Also provided are screening methods for identifying novel therapeutics that inhibit the MAP kinase-DEF domain interaction. This invention provides several advantages over known therapies that directly target the MAP

kinase signaling cascade. Typically, most compounds that inhibit the MAP kinase pathway are non-specific and inhibit more than one enzyme. Also, the targeted kinases, if effectively inhibited, are not available to perform normal physiological functions necessary for cell survival, resulting in toxicity to healthy as well as diseased cells. By contrast, the therapeutic methods of the present invention inhibit the activation of particular target proteins, leaving the MAP kinases enzymatically active and available to phosphorylate other, non-DEF domain-containing proteins. Diseased cells (e.g., cancerous cells) are often more susceptible to therapy because of the higher concentration of target protein, improving the likelihood of success for this approach.

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The principles of the invention are exemplified using the immediate early gene, c-Fos, but is not intended to be limiting. c-Fos functions as a molecular sensor for the duration of extracellular-signal-regulated kinase 1/2 (ERK1/2) signaling. c-Fos is known to be phosphorylated by ERK1/2 and RSK, resulting in increased stability of the protein. Therefore, the biological function of c-Fos differs under conditions where ERK1/2 signaling is sustained, rather than transient. Signaling is transduced by ERK1/2 binding to the DEF domain of c-Fos. Mutating the DEF domain inhibits c-Fos-mediated signaling and, ultimately, the downstream effects of ERK1/2 activation.

Further, Fos, Myc and Jun family proteins are transcription factors encoded by immediate early protooncogenes. Family members c-Fos, Fra-1, Fra-2, c-Myc, N-Myc, JunD, and JunB are frequently found to be amplified or upregulated in human cancers. Sustained ERK1/2 signaling is required for cell proliferation and ERK1/2 docking to these proteins occurs only when signaling is sustained.

25 Docking controls the growth-promoting function of these transcription factors.

Accordingly, ERK1/2 docking inhibitors may be clinically useful drugs because they will likely to inhibit a specific branch of ERK1/2 signaling and would, therefore, be less toxic than general ERK1/2 inhibitors.

5 Sustained ERK1/2 Activation Correlates With S Phase Entry

Treatment of quiescent Swiss 3T3 fibroblasts with platelet-derived growth factor (PDGF) stimulated S phase entry; whereas, treatment with epidermal growth factor (EGF) did not (Figure 1A). The activation kinetics and amplitude of ERK1/2 and RSK, however, were almost identical following a 5-10 minute exposure to either PDGF or EGF (Figure 1B). In both cases, hyperphosphorylated 10 (active) ERK1/2 was localized to the nucleus (Figure 1C). In contrast to ERK1/2 and RSK activation kinetics, rates of inactivation were faster in cells treated with EGF compared to PDGF (Figure 1B). ERK1/2 signaling remained elevated for at least 240 minutes following PDGF exposure, but returned to basal levels within 30-45 minutes following EGF withdrawal (Figure 1B). Notably, the sustained 15 ERK1/2 activity elicited by PDGF treatment remained localized to the nucleus (Figures 1C and 9A), demonstrating a tight correlation with S phase entry. Further, c-Fos protein expression was prolonged in cells treated with PDGF compared to those treated with EGF (Figure 1D). This indicates that c-Fos becomes stabilized when ERK1/2 signaling is prolonged, but is unstable when 20 ERK1/2 signaling is transient. c-Fos expression was not affected by either the addition of cycloheximide to cells 75 minutes after PDGF treatment (Figure 9B) or the addition of actinomycin D 20 minutes after PDGF treatment. Thus, the differences in c-Fos expression between PDGF- and EGF-treated cells arises from post-translational control. This conclusion is further supported by studies showing 25 that the transcriptional induction of c-fos and other IEGs by various growth factors is completed within 30-45 minutes.

ERK1/2 and RSK coordinately phosphorylate the extreme C-terminus of c-Fos at Ser 374 and Ser 362, respectively (Figure 2A). Mutating these residues to aspartate (Fos-DD), which mimics phosphorylation, results in enhanced transformation of fibroblasts by comparison to c-Fos having Ser 362 and Ser 374 mutated to alanine (Fos-AA) (Okazaki et al., EMBO J., 14: 5048-5059, 1995; Chen et al., Proc. Natl. Acad. Sci. USA, 90: 10952-10956, 1993). Thus, increasing the stability of c-Fos is not the only manner in which this transcription factor can regulate cellular transformation. Fos-AA appears to be differentially regulated compared to Fos-DD.

We have discovered that the addition of serum to fibroblasts results in a large, λ phosphatase-sensitive electrophoretic mobility shift of Fos-DD, compared to Fos-AA (Figures 2B left, and 10A). This effect correlates with increased incorporation of ³²P-orthophosphate that was consistently two to threefold greater for Fos-DD than Fos-AA (Figure 2B, right). This demonstrates that
 phosphorylation of Ser 362 and Ser 374 prime c-Fos for additional growth factor-regulated phosphorylation. As Fos-DD has greater transforming potential than Fos-AA, the regulation of primed c-Fos is critical for promoting fibroblast proliferation.

Phosphorylation of Primed c-Fos is MEK-dependent

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NIH 3T3 cells transfected with different Fos proteins were treated with the MEK inhibitor UO126 (Favata et al., J. Biol. Chem., 273: 18623-18632, 1998) to determine if the mitogen-regulated phosphorylation of Fos-DD is mediated by the Raf/MEK/MAPK pathway. UO126 inhibited the growth factor-regulated mobility shift of Fos-WT and Fos-DD (Figure 2C) and ERK1/2 activation (Figure 2C, bottom), indicated that ERK1/2 or downstream signaling molecules regulated primed c-Fos. Identical observations were made using NIH 3T3 cells expressing a

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conditionally active form of B-Raf (Δ B-Raf-ER) and treating these cells with tamoxifen instead of EGF (Figure 10B). To determine whether ERK1/2 could phosphorylate primed c-Fos *in vitro*, we used different hexahistidine-Fos fusion proteins as substrates. ERK1/2 efficiently phosphorylated Fos-WT (Figure 2D). The phosphorylation of Fos-AA and primed c-Fos, Fos-EE (S362E/S374E), by ERK1/2 was also easily detected *in vitro*, but the phosphorylation of Fos-EE compared with Fos-AA was consistently greater (Figure 2d). These results

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demonstrate that ERK1/2 can phosphorylate sites in c-Fos other than Ser 374 and that this phosphorylation is enhanced after priming of the C terminus. In contrast to ERK1/2, phosphorylation of c-Fos by ERK5 *in vitro*, another UO126-sensitive proline-directed kinase, was not observed (Figure 10C).

An ERK1/2 Targeting Motif Promotes the Phosphorylation of Primed Fos

The preference of ERK1/2 for primed c-Fos (Fos-EE) over Fos-AA demonstrates that C-terminal phosphorylation exposes additional phosphorylation 15 sites and/or a motif that would increase the efficiency of phosphorylation at these sites. Examination of the c-Fos sequence identified a site in the C terminus that has similarity with the ERK1/2 targeting motif, FXFP (SEQ ID NO: 1), known as a DEF domain. In c-Fos, this motif is FTYP (Figure 3A; SEQ ID NO: 2). Mutating either Phe 343 or Tyr 345 to alanine dramatically inhibited the 20 phosphorylation of primed c-Fos (Fos-EE) by ERK1/2 in vitro (Figure 3A). ERK1/2-regulated phosphorylation of substrates that contain DEF domains can be competitively inhibited in vitro with a synthetic peptide encompassing the DEF domain found in ELK-1. This peptide (FQFP; SEQ ID NO: 3) inhibited the phosphorylation of primed c-Fos in a concentration-dependent manner (Figure 25 3B). By contrast, a peptide with a mutant DEF domain (AQAP; SEQ ID NO: 4) was less efficient in inhibiting ERK1/2-mediated phosphorylation of primed c-Fos.

The ELK-1 peptide was then engineered to contain the c-Fos FTYP DEF domain (SEQ ID NO: 2). This peptide also inhibited primed c-Fos phosphorylation, but a mutant version (ATYP; SEQ ID NO: 5) did not (Figure 3C). In both cases, the IC₅₀ for the FQFP (SEQ ID NO: 3) and FTYP (SEQ ID NO: 2)peptides was approximately 80 μ M. The EGF-stimulated mobility shift of Fos-DD *in vivo* was also inhibited when Phe 343 or Tyr 345 were mutated to alanine (Figure 3D). These results demonstrate that the initial phosphorylation of c-Fos by ERK1/2 and RSK as the extreme C terminus expose an ERK1/2 docking site that allows ERK1/2 to phosphorylate additional sites.

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Based on this model, mutation of Phe 343 or Tyr 345 to alanine should prevent hyperphosphorylation of Fos-WT and not interfere with the priming phosphorylations, which are C-terminal to the DEF domain. Indeed, these mutations prevented the appearance of the slowest mobility, but still allowed a shift to the intermediate mobility (Figure 3E). This indicates that the DEF domain is not involved in directing ERK1/2 to prime c-Fos through Ser 374 phosphorylation. Instead, ERK1/2 docking through the DEF domain results in the hyperphosphorylation of primed c-Fos.

ERK1/2 phosphorylates Thr 325 and Thr 331 in primed c-Fos

There are two proline-directed threonine residues (Thr 325 and Thr 331) amino-terminal to the DEF domain in c-Fos (Figure 4A). Mutation of Thr 325 to alanine almost completely inhibited the phosphorylation of Fos-EE by ERK1/2 in vitro, and the additional mutation of Thr 331 to alanine was required to reduce the phosphorylation to background levels (Figure 4A). Thr 325 and Thr 331 were also phosphorylated in primed c-Fos (Fos-DD) in vivo, as evidenced by the complete loss of the mobility shift in the T325A/T331A mutant (Figure 4B). Individual mutation of Thr 325 or Thr 331 to alanine in the context of Fos-DD only partially

inhibited growth factor-regulated phosphorylation. Substituting alanines for Thr 325 and Thr 331 in the context of Fos-WT prevented the EGF-stimulated to the slowest mobility (Figure 4C). However, EGF treatment was associated with the appearance of the intermediate mobility form, resulting from priming

phosphorylation of ERK1/2 and RSK. Collectively, these observation demonstrate an ordered phosphorylation process whereby the initial phosphorylation of c-Fos at Ser 374 and Ser 362 (priming) exposes a DEF domain that mediates the hyperphosphorylation of c-Fos at Thr 325 and Thr 331. Further, Ser 374 phosphorylation is not regulated by docking. This is consistent with the phosphorylation of amino acids N-terminal, but not C-terminal, to DEF domains.

A phosphorylation-specific antiserum for Thr 325 in c-Fos (Figure 5A) was generated to investigate the mitogen-regulated phosphorylation of this residue in primed c-Fos. The antiserum showed little or no reactivity with Fos-WT or Fos-DD expressed in quiescent cells (Figure 5B, -EGF). However, after treatment with EGF, strong reactivity was associated with Fos-WT and Fos-DD, but not with Fos^{T325A} or Fos-DD^{T325A} (Figure 5B, +EGF). Priming of the extreme C terminus by ERK1/2 and RSK promotes additional phosphorylation of c-Fos *in vivo* (Figure 2B). To determine if this is caused by increased phosphorylation of Thr 325, Fos-WT and Fos-AA were expressed to similar levels in the ΔB-Raf-ER NIH 3T3 cells that were then treated with tamoxifen for varying times (Figure 5C). The phosphorylation of Thr 325 was greater in cells transfected with Fos-WT than those transfected with Fos-AA.

Mutating Phe 343 or Tyr 345 to alanine prevented the hyperphosphorylation of primed c-Fos (Figure 3). Specifically, the regulation of Thr 325 phosphorylation *in vivo* was inhibited when Phe 343 or Tyr 345 were mutated to alanine, either in the context of Fos-WT (Figure 5D) or Fos-DD (Figure 5E).

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In this later experiment, there is a strong correlation between the mobility shift of Fos-DD and increased Thr 325 phosphorylation confirming that the DEF domain in c-Fos increases the efficiency of Thr 325 phosphorylation *in vivo*.

The phosphorylation of Thr 325 is differentially regulated by ERK1/2-signal duration

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As shown above, the induction kinetics of c-Fos expression 30-45 min after addition of PDGF or EGF to Swiss 3T3 cells were identical (Figure 1D). This is consistent with a model in which an initial activation of ERK1/2 or RSK is sufficient for induction of c-fos IEG expression. To determine if ERK1/2 signal duration differentially regulates the phosphorylation of Thr 325 in endogenous c-Fos, we prepared extracts from Swiss 3T3 cells treated with EGF or PDGF for different times. Importantly, although c-Fos is present in cells after 45 or 60 min of EGF treatment, Thr 325 phosphorylation was not observed (Figure 6A). This is consistent with inactivation of ERK1/2 occurring before c-Fos is present (Figure 6A). By contrast, phosphorylation of Thr 325 increased 45-60 min after addition of PDGF (Figure 6B). Maximal Thr 325 phosphorylation persisted for at least 120 min (Figure 6B), but was still detected after 240 min. In Rat-1 fibroblasts, treatment with 100 µM lysophosphatidic acid (LPA) results in sustained ERK1/2 activity and S phase entry, whereas treatment with 0.1-1 µM LPA transiently activates ERK1/2 and no cell cycle progression occurrs. Although treatment of quiescent Rat-1 fibroblasts with mitogenic (100 µM) and no-mitogenic (0.5 µM) concentrations of LPA resulted in a similar induction of c-fos, phosphorylation of Thr 325 only occurred with 100 µM LPA (Figure 11). These findings correlate with the generation of transient and sustained ERK1/2 responses by 0.5 μM and 100 μM LPA, respectively (Figure 11).

Thus, differential phosphorylation of c-Fos occurs in different cell types and in response to agonists that directly activate tyrosine kinase receptors or heterotrimeric G protein-coupled receptors.

To show that the sustained phase of ERK1/2 signaling was required to

mediate the stabilization and hyperphosphorylation of endogenous c-Fos in Swiss
3T3 cells, ERK1/2 and RSK activity was inhibited by adding UO126 to cells that
had been treated with PDGF for 60 min (Figure 6D). Under this condition, the
phosphorylation of Thr 325 was completely inhibited and the electrophoretic
mobility of c-Fos increased (Figure 6C). Importantly, these UO126-induced
changes in the biochemical properties of c-Fos also preceded the rapid
disappearance of c-Fos protein. This result is consistent with hypophosphorylated
c-Fos being unstable. These observation show that the phosphorylation of c-Fos at
Thr 325 is tightly correlated with the activation/inactivation kinetics of ERK1/2 in
different cell types of provide clear evidence that c-Fos can function as sensor for
ERK1/2 signal duration.

The DEF domain and Thr 325/Thr 331 phosphorylation modulates c-Fos function

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In Hela cells expressing Fos-WT, AP-1 transcription factor activity was consistently three to fourfold above background levels (Figure 7A). Mutation of Thr 325 and Thr 331 to alanine reduced AP-1 activity by about 20%; whereas, mutating the DEF domain (F343A) reduced Fos-WT activity by about 65% (Figure 7A). These observations indicate that docking of ERK1/2 to c-Fos is important in regulating c-Fos transcriptional activity under conditions of growth factor stimulation. To determine if ERK1/2 docking to c-Fos can contribute to c-Fos function independently of the phosphorylation-mediated stabilization, Fos-WT, Fos^{T325A/T331A} or Fos^{Y345A} (a DEF domain mutant) were stably expressed in 208F

fibroblasts. The expression of the different Fos proteins (Figure 7B, bottom six panels) was equivalent to the level of endogenous c-Fos in serum-stimulated vector-infected cells (Figure 7B, top two panels) and was also localized to the nucleus. Western analysis of c-Fos expression in the quiescent cell lines also showed that they were expressed to similar levels (Figure 7C). The stable expression of Fos-WT promoted anchorage-independent growth in soft agar (Figure 7D), as expected. Mutating Thr 325 and Thr 331 to alanine significantly reduced the growth of 208F cells in soft agar suggesting that phosphorylation of these residues promotes cellular transformation (Figure 7D). However, replacing Thr 325 and Thr 331 with Asp enhanced c-Fos-mediated focus formation (Figure 12). Further, mutating the c-Fos DEF domain (Fos Y345A) completely inhibited the ability of c-Fos to transform 208F cells; more so than the c-Fos^{T325A/T331A} mutant (Figure 7D). These results demonstrate that ERK 1/2 docking to c-Fos contributes to transformation through mechanisms in addition to Thr 325/Thr 331 phosphorylation and that stabilization of c-Fos is not the only factor that regulates c-Fos function, as all proteins were expressed equally. In addition, it also demonstrates that ERK1/2 docking to c-Fos regulates biological activity.

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Mutating the DEF domain binding site inhibits target residue phosphorylation

In order to confirm the criticality of ERK1/2 docking with the DEF domain to phosphorylation of a target residue, the DEF domain binding site in ERK1/2 was mutated. Six single mutations in ERK1/2 (L198A, Y231A, L232A, L235A, Y261A, and D319N) were expressed in NIH 3T3. Several of these mutations have been previously described as forming the DEF domain binding site in ERK1/2 (Lee et al., Molec. Cell, 14: 43-55, 2004). Each ERK1/2 mutant was tagged with HA for later detection. EGF-stimulated kinase activity of wildtype and mutant

ERK1/2 was measured using RSK, a non-DEF domain-containing target protein, and c-Fos, DEF domain-containing target protein, as substrates in a standard ³²P-phosphorylation assay. Fos phosphorylation by ERK1/2 mutants with disrupted DEF domain binding pockets (L198A, Y231A, L232A, L235A, and Y261A) was almost completely absent (Figure 26). By contrast, RSK phosphorylation—a event that does not require ERK1/2 interaction with a DEF domain—was only moderately reduced (Figure 26).

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These results demonstrate that DEF domain binding is required for a target residue phosphorylation but that a disruption of the DEF domain inhibition does not abolish all kinase activity. Thus, the DEF domain binding event is separate and distinct from the phosphorylation event. Disruption of DEF domain binding may be used to selectively inhibit phosphorylation of a target protein, without significantly inhibiting the phosphorylation of non-target proteins (i.e., proteins that do not contain a DEF domain) by the same kinase. Thus, small molecule inhibitors and polypeptide inhibitors (e.g., naked DEF domains) which specifically inhibit DEF domain binding are useful for selectively inhibiting the phosphorylation of target proteins without causing the adverse effects associated with complete inhibition of a target kinase (e.g., a MAP kinase).

20 Mechanism of IEG Activation Through DEF Domain Binding

The mechanism described here employs an IEG product, typified by c-Fos, which functions as a molecular sensor that differentiates between differences in ERK1/2 and RSK signal duration, as well as their cytoplasmic/nuclear distribution (Figure 8). As observed for a large number of IEGs, the *c-fos* gene is transcriptionally induced within minutes of growth factor stimulation and therefore occurs with kinetics that are independent of differences in signal duration. Newly synthesized c-Fos protein has a half-life of about 30-45 min but, when

phosphorylated by ERK1/2 and RSK, the half-life is extended to at least 2h. Thus, when ERK1/2 is rapidly activated (transient signal), c-Fos is present in the nucleus, but is not phosphorylated, and is therefore unstable and degraded (Figure 8). By contrast, delayed inactivation of ERK1/2 (sustained signal) results in the efficient phosphorylation of c-Fos at its extreme C terminus, resulting in its stabilization for several hours. The initial priming phosphorylation in the C-terminus exposes a DEF domain that promotes additional ERK1/2-mediated phosphorylation events, increasing the efficiency of ERK1/2-regulated phosphorylation when ERK1/2 is only sub-maximally active (0.5-4 h after stimulation). Further, when priming and docking are inhibited by point mutation (Fos-AA and Fos^{F343A} or Fos^{Y345A}, respectively) ERK1/2 and/or RSK signals are unable to alter c-Fos function. These non-phosphorylable c-Fos mutants likely resemble the hypophosphorylated form of c-Fos that is present when ERK1/2 is rapidly inactivated during transient signaling and cells do not enter S phase.

Simply prolonging the half-life of c-Fos will not affect its role in promoting transformation. Instead, the combination of protein stabilization and DEF-mediated regulation allows c-Fos to function as sensor for ERK1/2. If c-Fos is not stabilized during the sustained phase of signaling, ERK1/2 will not target the c-Fos DEF domain. Therefore, stabilizing the IEG product is a critical first step if it is to function as a sensor for sustained ERK1/2 signals. The physiological importance of the c-Fos DEF domain is underscored by the fact that mutations in the DEF domain significantly reduced AP-1 activity and inhibit the transforming activity associated with wild-type c-Fos. However, the effect of mutating the DEF domain is stronger than the effect of mutating the phosphorylation sites that are controlled by this docking site, indicating that the DEF domain can have more than one action. An additional function of the DEF domain ERK1/2-mediated *trans*-phosphorylation of AP-1 complex proteins.

We described a general mechanism for cellular sensing of ERK1/2 signal strength and timing involving the FTYP (SEQ ID NO: 2) DEF domain present in many IEGs. Putative DEF domains are found in additional AP-1 proteins, such as Fra-1, Fra-2, Jun-B and JunD (Table 1). The proto-oncogene products c-Myc and N-Myc also contain putative DEF domains. The IEG product Egr-1 has a DEF domain and several putative proline-directed phosphorylation sites N-terminal to this domain that could enable Egr-1 to sense sustained signaling in PC12 cells and promote neuronal differentiation. In common with the c-Fos DEF domain, the other DEF domains highlighted in Table 1 show subtle deviation form the FXFP consensus (SEQ ID NO: 1), with respect to the presence of phenylalanine at positions 1 and 3 indicating that tyrosine can be tolerated at either site.

Table 1. DEF Domains in Immediate Early Gene Products		
IEG	Amino Acid Sequence	SEQ ID NO.
c-Fos	314-GPMVTELEPLCTP-VVTCTPSCTTYTSSFVFTYPEEADS	6
Fra-1	²¹¹ -GP-VLEPEALHTPTLMT-TPSLTPFTPSLVFT YP STPEP	7
Fra-2	¹⁶⁹ -GGFYGE-EPLHTP-IVVTSTPAITPGTSNLV FTYP SVLEQ	8
Fos-B	²⁸³ -HSEVQV-LGDPFPVV-SPS-YTSSFVLTCPEVSAF	9
JunD	⁸⁷ -LLASPDLGLLKLASPELERLIIQS-NGLVTTTPTST-Q FLYP KV	10
JunB	⁷¹ -GQGSDTGASLKLASSELERLIVPNSNGVITTTPTPPGQ YFYP RG	11
c-Jun	60-LLTSPDVGLLKLASPELERLIIQSSNGHITTTPTPT-QFLCPKN	12
с-Мус	⁵⁶ -LPTPPLSPSRRSGLCSPSYV	13
	¹⁸¹ -LTA-AASECIDPSVV FPYP LND	14
N-Myc	⁷⁷ -AQSPGAGAASPAGRGHGGAAGA	15
	110-AHPAAECVDPAVVF PFP VNK	16

Egr-1	184-QSPPLSCAVPSNDSSPIYSAAPTFPTPNTD	17
	²⁴⁷ -PMIPDYLFPQQ	18
MPer1	⁶⁹⁷ -PRGGPQPLPPAPTSVPPAAFPAPLVTPMVALPN YLFP TPSSY	19

DEF domains are in bold and number indicate amino acid position. Sequences are from rat (c-Fos, Fra-1, and Fra-2), mouse (FosB, JunD, c-Jun, c-Myc, and Egr-1), or human (JunB, N-Myc, and mPer1).

Screening Methods to Identify Inhibitors of DEF Domain Binding

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DEF Domain Binding Assessment using a Phospho-specific Antibody We have developed a cell-based assay which is used to screen small molecule compound libraries (Figure 25). In this assay, rat 208F fibroblasts that stably express c-Fos are cultured in a 384-well plate and deprived of serum growth factors for 24 hours. Cells are then treated with EGF for 15 minutes and fixed with 3.7% formaldehyde. Permeabilized fixed cells are incubated with DAPI to stain the nuclei and an anti-phospho-ERK1/2 mouse monoclonal antibody and an anti-phospho-T325 Fos rabbit polyclonal antibody for 2 hours. Anti-mouse Alexa594-conjugated IgG and an anti-rabbit FITC-conjugated IgG are added to each well and unbound antibody is removed by several washes. The fluorescence intensity of both fluorophores in each well can be detected using an automated epifluorescence microscope or Autoscope (Universal Imaging Systems, Inc.). A clear increase in T325 phosphorylation was observed in cells treated with EGF (indicating that ERK1/2 docking to Fos has taken place). In the same population of cells, the phosphorylation of ERK1/2 also increased, thus indicating its activation by EGF. Under these conditions, only background levels of fluorescence were detected when both phospho-specific antibodies were omitted, and the secondary antibodies show no cross-species reactivity.

Inhibition of ERK1/2 docking to the c-Fos DEF domain in vivo could result from compounds that (a) are generally toxic, (b) prevent the activation of ERK1/2, (c) prevent the growth factor regulated translocation of ERK1/2 into the nucleus where c-Fos is localized or (d) directly antagonize ERK1/2 docking to the DEF domain. The assay we have developed naturally excludes the first three possibilities. First, toxicity will be reflected by nuclear integrity as visualized with DAPI staining. Second, inhibition of ERK1/2 activation/activity will be apparent from the phospho-ERK1/2 fluorescence signal. Third, nuclear translocation of ERK1/2 can be verified by manually examining images from wells that show 10 decreased c-Fos phosphorylation. Therefore, candidate compounds that specifically inhibit ERK1/2 binding to the DEF domain are defined as compounds that decrease the phosphorylation of T325 in c-Fos but which have no effect on ERK1/2 activation or its localization. Although this assay is exemplified using rat fibroblasts, it may be performed using any appropriate cell type including, for example, myoblasts, epithelial cells, and hepatocytes. 15

Interaction-Trap Assays

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A standard yeast two-hybrid assay may be used to assess the effect of a test compound on the MAP kinase-DEF domain interaction (Mendelsohn and Brent, Curr. Opin. Biotechnol. 5:482-486, 1994). Typically, a vector encoding a synthetic or 20 naturally occurring peptide containing a DEF domain, covalently bound a DNA binding domain (e.g., GAL4), is transfected into yeast cells containing a reporter gene operably linked to a binding site for the DNA binding domain. Further, a vector encoding either the native MAP kinase of interest, or a synthetic fragment 25 containing the sequence that interacts with the target DEF domain, covalently bound to a transcriptional activator (e.g., GalAD) is also transfected.

The effectiveness of a test compound is then assessed by growing the yeast in the presence of the compound and measuring the level of reporter gene expression.

GST Pulldown Assays

5 The interaction of a MAP kinase with a DEF domain may be examined using a GST-fusion protein binding study. A vector encoding a naturallyoccurring or synthetic polypeptide containing the DEF domain of interest is fused to GST and expressed in a host cell (e.g., E. coli or Saccharomyces spp.). The GST fusion protein is then contacted with a MAP kinase in the presence and 10 absence of a test compound. The MAP kinase may be naturally expressed by the host cell or may be expressed from a second vector inserted into the host cell. Following incubation with the test compound, the host cells are lysed and the GST fusion proteins are recovered using glutathione-Sepharose (GSH-Seph) beads. Typically, the GST fusion proteins are released from the GSH-Seph by boiling and 15 the proteins visualized by electrophoretic separation on an SDS-PAGE gel. A skilled artisan will readily understand that the GST-Pulldown assay described here can be readily adapted to a cell-free assay by incubating the purified GST fusion protein with a purified recombinant MAP kinase.

Fluorescence Polarization Assay

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A variety of well known cell-free techniques may be used to assess the effects of a test compound on the interaction between a MAP kinase and a DEF domain-containing target protein. Fluorescence polarization assays are particularly useful for this purpose. In this assay, a peptide (about 6-12 amino acids) containing a DEF domain at its C-terminus and a fluorophore (e.g., fluorescein) conjugated to its N-terminus is incubated in the presence and absence of increasing amounts of recombinant MAP kinase (e.g., GST-ERK1; 0.01-1 µM) for 10

minutes at room temperature. Aliquots from each reaction are placed in a plate black-walled microtiter (e.g., 384-well) plate and polarization measured using an Analyst plate reader. Increasing concentrations of the MAP kinase causes an increase in polarization. Titrating in the "free" DEF domain-containing peptide (i.e., un-conjugated) inhibits the change in polarization, whereas the mutated DEF domain peptide does not. The appearance of low polarization, even in the presence of high concentrations of kinase, indicates flexible binding of the DEF domain to the kinase and suggests the presence of the propeller effect. Designing shorter dye-conjugated DEF domain-containing peptides usually alleviates this problem. The effect of standard assay variables, including incubation time, temperature, pH (7.2-8.5), and buffers, on polarization is readily controlled during routine assay optimization.

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This assay is readily adaptable for identifying test compounds that inhibit binding of a MAP kinase to a DEF domain. The use of automated liquid handling systems and plate readers makes this assay readily adaptable to a high-throughput format for screening large numbers of test compounds. For compound screening, the test compound is added to a mixture of the fluorescently labeled DEF domain-containing peptide and the target MAP kinase. Compounds that inhibit the polarization increase (or cause a decrease in polarization) resulting from increasing amounts of the MAP kinase are therapeutic candidates.

Identification of Test Compounds as Potential Therapeutics

We have identified a variety of DEF domain-containing target proteins that have been implicated in a variety of diseases. The particular DEF domain or target protein may be substituted for c-Fos in any of the exemplary assays described here. Further, the lists of target proteins provided are not intended to be limiting. Other target proteins are easily identified based on the availability of a DEF domain.

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Test compounds having antineoplastic activity are those that inhibit binding of a MAP kinase (e.g., ERK 1/2) to the DEF domain of any of the proteins of Table 1 (except mPer1) or Table 2. Test compounds that are useful for treating cardiovascular disorders inhibit MAP kinase binding to the DEF domain of the proteins identified in Table 3. Test compounds that are useful for treating acute and chronic inflammation or inflammatory disorders inhibit MAP kinase binding to the DEF domain of the proteins identified in Table 4. Test compounds that are useful for treating a variety of metabolic disorders inhibit MAP kinase binding to the DEF domain of the proteins identified in Table 5. Test compounds that are useful for treating a variety of nervous system disorders (e.g., central and peripheral neuropathies) and behavioral disorders (e.g., psychosis, schizophrenia, autism, Down's Syndrome, Parkinson's Disease, Alzheimer's Disease, epilepsy, Cockayne syndrome, depression, and opiate addiction) inhibit MAP kinase binding to the DEF domain of the proteins identified in Table 6. Test compounds useful for treating sleep disorders inhibit MAP kinase binding to the DEF domain of the Per proteins (Table 1: mPer; Table 7). Other potentially useful therapeutics inhibit MAP kinase binding to the DEF domain of the PKA-anchoring proteins (AKAPs) (Table 7). In each of Tables 2-7, the alpha-numeric Accession Codes refer to the SWISS-PROT accession numbers. The numeric Accession Codes refer to the GENPEPT accession numbers. In each case, the DEF domain is underscored.

Administration of a DEF Domain Inhibitors or Candidate Compounds for the Treatment of Disease

As described above, ERK1/2 activate several IEG products through an interaction with the DEF domain and a subsequent phosphorylation event. It is also well known that activation of certain IEGs, and the proteins identified in

Table 2, cause cellular proliferation and may cause tumor promotion and progression. Accordingly, this invention also provides methods and compositions for antineoplastic (i.e., cancer) therapy by administering DEF domain inhibitors. Likewise, therapy for cardiovascular disorders, inflammatory disorders, metabolic disorders, neuropathies and behavioral disorders, and sleep disorders may be provided by inhibiting MAP kinase binding to the DEF domain of one or more of the proteins identified in Table 3, 4, 5, 6, and 7, respectively. Useful DEF domain inhibitors include compounds that bind to the DEF domain of target proteins and prevent the binding of the target kinases. Also, DEF domain inhibitors include "bait" proteins that bind activated target kinases but do not cause cellular proliferation or tumor promotion and/or progression.

In addition to candidate compounds identified using the screening methods of this invention, DEF domain inhibitors can be created by inserting, by artifice, a DEF domain into a non-target protein. The cellular activation/proliferation pathway described herein is limited by the presence of activated target kinase, not by the availability of target proteins. Thus, a DEF domain that is present in a non-target protein effectively "baits" the target kinase, reducing its availability to phosphorylate the target proteins. DEF domains suitable for therapy have the general structure: F/Y—X₁—F/Y—X₂ (SEQ ID NO: 28). Desirably, X₂ is proline. Most desirably, the DEF domain is identical to the DEF domain of the target protein to which therapy is directed. For example, Figures 3B and C demonstrate that the "naked" DEF domains FQFP (SEQ ID NO: 3) and FTYP (SEQ ID NO: 2) are effective inhibitors of target protein phosphorylation. Substitution of phenylalanine for alanine in these polypeptides results in approximately a two-fold reduction in potency.

Accordingly, therapy can be provided by administering pharmaceutical formulations containing a naked DEF domain. Typically, these polypeptides are administered by parenteral injection such as intravenous, intramuscular, or subcutaneous injection. These small polypeptides may be administered in any appropriate formulation including, for example, in a liposomal formulation. The polypeptides may also be injected directly into a solid tumor.

Alternatively, therapy can be achieved by administering a chimeric protein consisting of a DEF domain that is engineered into a non-target protein. Typically, the chimeric protein will "display" the four amino acid DEF domain on a hydrophilic face, making it available to bind to the activated target kinase. The non-target protein can be chosen based upon the desired pharmacokinetic or pharmacodynamic effect and is readily determined by a person of ordinary skill. For example, a DEF domain inhibitor sequence may be engineered into a serum protein such as albumin or ceruloplasmin in order to prolong the plasma half life. Alternatively, the DEF domain may be engineered into a protein that promotes uptake into a particular cell type.

Pharmaceutical Formulations

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The peptide agents and candidate compounds of the invention can be administered to a subject, e.g., a human, directly or in combination with any pharmaceutically acceptable carrier or salt known in the art. Pharmaceutically acceptable salts may include non-toxic acid addition salts or metal complexes that are commonly used in the pharmaceutical industry. Examples of acid addition salts include organic acids such as acetic, lactic, pamoic, maleic, citric, malic, ascorbic, succinic, benzoic, palmitic, suberic, salicylic, tartaric, methanesulfonic, toluenesulfonic, or trifluoroacetic acids or the like; polymeric acids such as tannic acid, carboxymethyl cellulose, or the like; and inorganic acids such as hydrochloric

acid, hydrobromic acid, sulfuric acid phosphoric acid, or the like. Metal complexes include zinc, iron, and the like. One exemplary pharmaceutically acceptable carrier is physiological saline. Other physiologically acceptable carriers and their formulations are known to one skilled in the art and described, for example, in Remington's Pharmaceutical Sciences, (19th edition), ed. A. Gennaro, 1995, Mack Publishing Company, Easton, PA.

Pharmaceutical formulations of a therapeutically effective amount of a peptide agent or candidate compound of the invention, or pharmaceutically acceptable salt-thereof, can be administered orally, parenterally (e.g. intramuscular, intraperitoneal, intravenous, or subcutaneous injection), or by intrathecal or intracerebroventricular injection in an admixture with a pharmaceutically acceptable carrier adapted for the route of administration.

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Methods well known in the art for making formulations are found, for example, in Remington's Pharmaceutical Sciences (19th edition), ed. A. Gennaro, 1995, Mack Publishing Company, Easton, PA. Compositions intended for oral use may be prepared in solid or liquid forms according to any method known to the art for the manufacture of pharmaceutical compositions. The compositions may optionally contain sweetening, flavoring, coloring, perfuming, and/or preserving agents in order to provide a more palatable preparation. Solid dosage forms for oral administration include capsules, tablets, pills, powders, and granules. In such solid forms, the active compound is admixed with at least one inert pharmaceutically acceptable carrier or excipient. These may include, for example, inert diluents, such as calcium carbonate, sodium carbonate, lactose, sucrose, starch, calcium phosphate, sodium phosphate, or kaolin. Binding agents, buffering agents, and/or lubricating agents (e.g., magnesium stearate) may also be used. Tablets and pills can additionally be prepared with enteric coatings.

Liquid dosage forms for oral administration include pharmaceutically acceptable emulsions, solutions, suspensions, syrups, and soft gelatin capsules. These forms contain inert diluents commonly used in the art, such as water or an oil medium. Besides such inert diluents, compositions can also include adjuvants, such as wetting agents, emulsifying agents, and suspending agents.

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Formulations for parenteral administration (i.e., intravenous, intramuscular, and subcutaneous injection) include sterile aqueous or non-aqueous solutions, suspensions, or emulsions. Examples of suitable vehicles include propylene glycol, polyethylene glycol, vegetable oils, gelatin, hydrogenated naphalenes, and injectable organic esters, such as ethyl oleate. Such formulations may also contain adjuvants, such as preserving, wetting, emulsifying, and dispersing agents. Biocompatible, biodegradable lactide polymer, lactide/glycolide copolymer, or polyoxyethylene-polyoxypropylene copolymers may be used to control the release of the compounds. Other potentially useful parenteral delivery systems for the proteins of the invention include ethylene-vinyl acetate copolymer particles, osmotic pumps, implantable infusion systems, and liposomes.

Liquid formulations can be sterilized by, for example, filtration through a bacteria-retaining filter, by incorporating sterilizing agents into the compositions, or by irradiating or heating the compositions. Alternatively, they can also be manufactured in the form of sterile, solid compositions which can be dissolved in sterile water or some other sterile injectable medium immediately before use.

The amount of active ingredient in the compositions of the invention can be varied. One skilled in the art will appreciate that the exact individual dosages may be adjusted somewhat depending upon a variety of factors, including the protein being administered, the time of administration, the route of administration, the nature of the formulation, the rate of excretion, the nature of the subject's conditions, and the age, weight, health, and gender of the patient. Generally,

dosage levels of between 0.1 µg/kg to 100 mg/kg of body weight are administered daily as a single dose or divided into multiple doses. Desirably, the general dosage range is between 250 µg/kg to 5.0 mg/kg of body weight per day. Wide variations in the needed dosage are to be expected in view of the differing efficiencies of the various routes of administration. For instance, oral administration generally would be expected to require higher dosage levels than administration by intravenous injection. Variations in these dosage levels can be adjusted using standard empirical routines for optimization, which are well known in the art. In general, the precise therapeutically effective dosage will be determined by the attending physician in consideration of the above identified factors.

The protein or candidate compound of the invention can be administered in a sustained release composition, such as those described in, for example, U.S. Patent No. 5,672,659 and U.S. Patent No. 5,595,760. The use of immediate or sustained release compositions depends on the type of condition being treated and the desired pharmacokinetic profile. For preventive or long-term treatments, a sustained released composition may be preferred.

The protein or candidate compound of the present invention can be prepared in any suitable manner. The protein or candidate compound can be isolated from naturally occurring sources, recombinantly produced, or produced synthetically, or produced by a combination of these methods. The synthesis of short peptides is well known in the art. See e.g. Stewart et al., Solid Phase Peptide Synthesis (Pierce Chemical Co., 2d ed., 1984).

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Methods

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Cell Culture

NIH 3T3 fibroblasts were transfected with Lipofectamine (Invitrogen, Carlsbad, CA) and then cultured for 18 h in DMEM/10% calf serum. Swiss 3T3 fibroblasts expressing a conditionally active form of B-Raf, were cultured in DMEM containing 10% fetal bovine serum (FBS). Before stimulation, cells were cultured in DMEM containing 20 mM HEPES (starving medium) for 24 h (NIH 3T3 or 208F) or 48 h (Swiss 3T3). Rat-1 fibroblasts were cultured for 48 h, washed with starving medium and culture for an additional 24 h in starving medium. For AP-1 assays, Hela cells were transfected with Lipofectamine for 6 h and then cultured for an additional 16 h before cell lysis and assay of luciferase activity (Promega). EGF and PDGF (Invitrogen) were reconstituted in sterile water containing 0.1% BSA. LPA (Aventi Polar Lipids, Alabaster, AL) was reconstituted in 50% ethanol before sonication for 30 min.

Retroviruses used to infect rat 208F cells were produced as described previously (Chen et al., Oncogene, 12:1493-1502, 1996). Neomycin-resistant pools of c-Fos-expressing cells were assayed for anchorage-independent growth or focus formation. Metabolic labeling with ³⁵S-methionine or ³²P-orthophosphate (performed in parallel) was performed as described by Chen et al.

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Cell lysis and western analysis

Cell extracts were prepared as described previously (Richards, et al., Curr. Biol., 9: 810-820, 1999). To analyze shifts in c-Fos mobility, samples were resolved on a 7.5% SDS-polyacrylamide gel electrophoresis (PAGE) gel, transferred to nitrocellulose and probed with anti-c-Fos antibody (Update Biotechnology Inc., Lake Placid, NY). This antibody is specific for c-Fos and does not cross-react with FosB, Fra-1 or Fra-2. For ERK1/2-MAPK western

analysis, a polyclonal anti-ERK 1/2 antibody to an anti-phospho-p42/p44 MAPK monoclonal antibody was used (Sigma, St. Louis, MO). Phosphatase treatment of cell extract was performed for 30 min on ice using λ protein phosphatase (New England Biolabs, Beverly, MA). To generate antiserum specific for phosphorylated Thr 325 in c-Fos, residues 317-329 (VTELEPLCTPVVT) (SEQ ID NO: 20) were synthesized (underlined residue is phospho-Thr at position 325), conjugated to keyhole limpet haemacyanin and injected into rabbits (Research Genetics, Inc., Huntsville, AL). To determine the specificity of the antiserum, extracts from cells expressing vector or c-Fos proteins were immobilized on nitrocellulose and probed with a solution of this anti-serum (1:3000) for 12 h at 4 °C.

Recombinant protein purification

M15pREP4 cells transformed with pDS56-(His)₆Fos or pETHis₆/ERK2 and
15 MEK 1 R4F were cultured at 25 C until an OD₆₀₀ of 0.7 was attained. Cells were
then incubated in the presence of 1mM isopropyl-β-D-thiogalactoside (IPTG) for
an additional 12 h at 25 ° C. and then harvested by centrifugation. Pellets were
resuspended in column buffer (20 mM Tris-HCl at pH 8.0, 200 mM sodium
chloride, 10% glycerol, and 10 mM imidazole) and cells were lysed by passage
20 through a French Press. The (His)₆ proteins were purified using Nickel-NTAagarose resin (Qiagen, Alencia, CA), dialyzed in column buffer containing 50%
glycerol and then stored at -20°C.

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In vitro kinase reactions

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Phosphorylation of (His)₆-Fos by ERK1 immunoprecipitated from NIH3T3 cells, activate (His)₆-ERK2 (ref. 42) or FLAG-ERK5/BMK1, was performed in kinase buffer containing 10 μ Ci γ^{32} P-ATP at 30 C (Chen *et al.*, *Mol. Cell. Biol.* 10:3204-3215, 1990). Endogenous ERK1 and RSK kinase activities were performed as described previously (Chung *et al.*, *Mol. Cell. Biol.*, 11:1868-1871, 1991). The HPLC-purified synthetic peptides used in the competition kinase assays were mixed with (His)₆-Fos-EE before addition of activated ERK2 and γ ³²P-ATP. The peptides derived from ELK-1 were as follows:

10 RRPRSPAKLSFQFPSFQFP (SEQ ID NO: 21); RRPRSPAKLSAQAPSAQAP (SEQ ID NO: 22); RRPRSPAKLSFTYPSFTYP (SEQ ID NO: 23); RRPRSPAKLSATYPSATYP (SEQ ID NO: 24).

Immunofluorescence

Swiss 3T3 cells (1.35 x 10⁵ per 35-mm dish) were cultured on poly-Llysine-coated glass coverslips for 24 h and serum-starved for 48 h. Cyclohexamide
(14 μg/ml) was delivered in dimethyl sulphoxide. After stimulation with growth
factors, cells were washed with ice-sold PBS containing 0.1% BSA, fixed with
3.7% formaldehyde for 10 min at room temperature and permeabilized with 0.2%
Triton-X100 for 5 min. Analysis of c-Fos expression was performed using a rabbit
anti-human c-Fos antibody (1:500, Upstate Biotechnology Inc.) under conditions
described by the manufacturer. Conditions for phospho-p24/p44 MAPK
immunofluorescence were identical to those used for c-Fos, except that a
monoclonal phospho-MAPK antibody was used (Sigma). Coverslips were
25 mounted in Citifluor (Ted Pella Inc., Redding, CA) and examined under
epifluorescent illumination.

Bromodeoxyuridine (BrdU) incorporation

Swiss 3T3 cells were cultured as described for immunofluorescence studies, treated with growth factors and 20 µM BrdU Labeling Reagent (Amersham Life Sceinces Inc., Piscataway, NJ) for 20 h at 37 °C. For immunofluorescence analysis, a mouse anti-BrdU monoclonal (Amersham Life Sciences Inc.) supplemented with DNAase I (Invitrogen) was used.

Other Embodiments

All publications and patent applications cited in this specification are herein incorporated by reference as if each individual publication or patent application were specifically and individually indicated to be incorporated by reference.

Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be readily apparent to those of ordinary skill in the art in light of the teachings of this invention that certain changes and modifications may be made thereto without departing from the spirit or scope of the appended claims.

What is claimed is:

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•	ABEE 2. ONCOLOGI	Amino
Accession Code		Acid Target Sequence
1 PIP3_HUMAN	1-phosphatidylinositol-4,5-bisphosphate phosphodiesterase beta 3 (EC 3.1.4.11) (Phosphoinositide phospholipase C) (PLC-beta-3) (Phospholipase C-beta-3). 26S proteasome non-ATPase regulatory subunit 1 (26S proteasome regulatory subunit S1) (26S	777 DEEP <u>FDFP</u> KWLPTL
2 PSD1_HUMAN	proteasome subunit p112). 26S proteasome non-ATPase regulatory subunit 1 (26S proteasome regulatory subunit S1) (26S	767 TQFW <u>FWFP</u> LSHFLSL
3 PSD1_HUMAN	proteasome subunit p112).	807 KPST <u>FAYP</u> APLEVPK
4 RT31_HUMAN 5 PDPK HUMAN	28S ribosomal protein S31, mitochondrial precursor (S31mt) (MRP-S31) (Imogen 38). 3-phosphoinositide dependent protein kinase-1 (EC 2.7.1.37) (hPDK1).	295 IKLEYDFPEKFFPKA
6 RS3_HUMAN	40S ribosomal protein S3.	73 VOKR <u>EGFP</u> EGSVELY
7 P52K HIMAN	52 kDa repressor of the inhibitor of the protein kinase (p58IPK- interacting protein) (58 kDa interferon-induced protein kinase- interacting protein) (P52rIPK) (Death associated protein 4).	18 DLAF <u>FRFP</u> RDPARCQ
8 B53A HUMAN	53 kDa BRG1-associated factor A (Actin-related protein Baf53a) (ArpNbeta).	283 PTVHYEFPNGYNCDF
9 AAK1_HUMAN	5'-AMP-activated protein kinase, catalytic alpha-1 chain (EC 2.7.1) (AMPK alpha-1 chain).	273 DLPKYLFPEDPSYSS
10 AAK2_HUMAN	5'-AMP-activated protein kinase	271 DLPS <u>YLFP</u> EDPSYDA 252 VETDYTEDI AEKYKA
11 RLAO_HUMAN		152 RRAKEKEPGROKIHI
	605 ribosomai protein ב זיט (עוש) 80 kda MCM3-associated profei	184 GLTPFSFPQVTNSSV
8 14 ASH3 MOUSE	-	
15 ASH3_HUMAN	-	69 EPCP <u>FSFP</u> MPYPNYR
16 AD10_HUMAN	disintegrin-metalloprotease) (Kuzbanian protein homolog).	286 PTNP <u>FRFP</u> NIGVEKF
17 AD12 HIMAN	ADAM 12 precursor (EC 3,4.24) (A disintegrin and metalloproteinase domain 12) (Meltrin alpha).	382 ASTGYPFPMVFSSCS
18 BS69 HUMAN	Adenovirus 5 E1A-binding protein (BS69 protein).	
19 CYA4_HUMAN	Adenylate cyclase, type IV (EC 4.6.1.1) (ATP pyrophosphate-lyase) (Adenylyl cyclase).	
20 AF4_HUMAN	AF-4 protein (Proto-oncogene AF4) (FEL protein). A kinase anchor protein 11 (Protein kinase A anchoring protein 11) (PRKA11) (A kinase anchor	381 EPSKEPFFIKUSORV
21 AK11 HUMAN	protein 220 kDa) (AKAP 220) (hAKAP220).	661 EVCQ <u>FSYP</u> QTPASPQ
I	A-kinase anchor protein 3 (Protein kinase A anchoring protein 3) (PRKA3) (A-kinase anchor protein 110 kDa) (AKAP 110) (Sperm oocyte binding protein) (Fibrousheathin I) (Fibrous sheath protein of	
22 AKA3_HUMAN	95 kDa) (FSP95).	490 SDIS <u>FEYP</u> EDIGNLS
144741111 7114 00	ALK tyrosine kinase receptor precuisor (EC 2.7.1.112) (Ariapiasuc tyripitorita nitrase) (CD240	264 I ECSEDEPCEI EYSP
23 ALK_HUMAN 24 ANR5 HUMAN	anugeri). Ankvrin repeat domain protein 5.	437 VIPE <u>YAFP</u> RRQDGGP
25 ATR_HUMAN	Anthrax toxin receptor precursor (Tumor endothelial marker 8).	421 PEQE <u>YEFP</u> EPRNLNN

193 DPEH <u>FPFP</u> APANAPL 65 LPLF <u>FTFP</u> PPGDTWE	362 RPSL <u>FVYP</u> EESLVIG 88 GITV <u>FYFP</u> RCQGFTS 240 SSGI <u>FQFP</u> LNLCTKT 483 KDVMFFFPFFFOLOTK	29 QIPK <u>YCFP</u> NYVGRPK 825 DTEG <u>EKYP</u> LGHEVNH	254 QDFS <u>YFFP</u> DDPPTFI 627 EEGP <u>FTFP</u> NGEAVEH	226 KAGA <u>YDFP</u> SPEWDIV	282 HIQL <u>FLFP</u> KSHDITQ 98 EALE <u>FYYP</u> EHFTLLT 86 ESLE <u>FYYP</u> ELYKLVT	323 QEFL <u>FVFP</u> FSCRQLQ 728 LENS <u>WLFP</u> TRIGGNF	202 LPRN <u>FHYP</u> PDGYSRH	162 PEGS <u>FQYP</u> ASYHSNQ 27 SSAA <u>FGFP</u> RGAGPAQ 102 GGGD <u>FDYP</u> GAPAGPG 84 DPRP <u>FAYP</u> PHTFGPD 673 SPAP <u>FPFP</u> EAPGSLP 1457 RSLI <u>YKFP</u> STFYEGR 241 SYAK <u>FLYP</u> TNALVTH
 A-Raf proto-oncogene serine/threonine-protein kinase (EC 2.7.1) (A-raf-1) (Proto-oncogene Pks). ATP-binding cassette, sub-family A, member 3 (ATP-binding cassette transporter 3) (ATP-binding cassette 3) (ABC-C transporter). ATP-dependent DNA helicase II, 70 kDa subunit (Lupus Ku autoantigen protein p70) (Ku70) (70 kDa subunit of Ku antigen) (Thyroid-lupus autoantigen) (TLAA) (CTC box binding factor 75 kDa subunit) 	ඉ	ဓ္တ	transcription factor-related chr transcription factor-related chr 55 calcium and DAG-regulated g Calcium/calmodulin-depender			Caspase recruitment domain protein 15 (Nod2 protein) (Inflammatory bowel disease protein 1). Caspase recruitment domain protein 6.	Catenin delta-1 (p120 catenin) (p120(ctn)) (Cadherin-associated Src substrate) (CAS) (p120(cas)). Catenin delta-2 (Delta-catenin) (Neural plakophilin-related ARM-repeat protein) (NPRAP)	(Neurojungin) (GT24). CCAAT/enhancer binding prote CCAAT/enhancer binding prote CCAAT/enhancer binding prote CCAAT/enhancer binding prote Cdc42 GTPase-activating prote Cdc42 guanine nucleotide exch Cdc5 and abl enzyme substrate
26 KRAA_HUMAN 27 ABC3_HUMAN	28 KU70_HUMAN 29 AXU1_HUMAN 30 1275113	31 1154493 32 ACTY_HUMAN 33 BRC1_HUMAN	34 BA1A_HUMAN 35 39288	36 KCCA_HUMAN	-37 CFLA_MOUSE 38 CARA_HUMAN 39 CARB_HUMAN	40 CARF_HUMAN 41 CAR6_HUMAN	42 CTD1_HUMAN	43 CTD2_HUMAN 44 CEBA_HUMAN 45 CEBA_HUMAN 46 CEBE_HUMAN 47 5020264 48 ZIZ1_HUMAN 49 CBL2_HUMAN

Clathrin coat assembly protein AP50 (Clathrin coat associated protein AP50) (Plasma membrane

or (Ceir recognition motecule Casprs). 128 128 Lhomolog) (hCm) (CGI-201) (MSTP021). 800	. homolog) (hCrn) (CGI-201) (MSTP021).	. nomolog) (ncm) (cGI-201) (MSTP021).	74	158	138 TRFS <u>YAFP</u> KEFPYRM	598 SSLNFSFPSLPTMGQ	Cytochrome P450 2A12 (EC 1.14.14.1) (CYPIIA12) (Steroid hormones 7- alpha-hydroxylase)	456 QNFR <u>FKFP</u> RKLEDIN	EC 2.7.1.112) (Bone marrow kinase BMX) (Epithelial and	3). 273 SKIS <u>WEFP</u> ESSSSEE	ospholipase A2 (EC 3.1.1.4) (Phosphatidylcholine		cleotidase cytosolic II).	sapiens] 229 PTNPEREPNISVEKF	DNA (cytosine-5)-methyltransferase 3A (EC 2.1.1.37) (Dnmt3a) (DNA methyltransferase HsallIA)	861 MERVEGEPVHYTDVS	DNA (cytosine-5)-methyltransferase 3B (EC 2.1.1.37) (Dnmt3b) (DNA methyltransferase HsalIIB)	805 LERIFGFPVHYTDVS	(DNA methyltransferase homolog		ein homolog 3).	389	7.7.7) (DNA polymerase II subunit B).	DNA polymerase gamma subunit 2, mitochondrial precursor (EC 2.7.7.7) (Mitochondrial DNA	(Foldstadia) (initrolia) (Unita polyliterase gaillitia accessory 33 KDa			.99.1.3). 1444 FGNLFSFPSYSQKSE fan homolog	<u>+</u>	Dual specificity mitogen-activated protein kinase kinase 4 (EC 2.7.1) (MAP kinase kinase 4) (JNK	activating kinase 1) (c-Jun N- terminal kinase kinase 1) (JNKK) (SAPK/ERK kinase 1) (SEK1).	Dual snecificity profein phosphatase 1 (FC 3 4 3 48) (FC 3 4 3 46) (MAP kinase phosphatase 4)	(MKP-1) (Protein-tyrosine phosphatase CL100) (Dual specificity protein phosphatase hVH1).	
rated protestr-III	Ork like protein	Ordering protein.		CID-binding SR-like protein RA4 (Fragment).	Cyclin C.	Cyclin T1 (Cyclin T) (CycT1).	Cytochrome P450 2A12 (EC 1.14.14.1) (CY	(Testosterone 7-alpha-hydroxylase).	Cytoplasmic tyrosine-protein kinase BMX (E	endothelial tyrosine kinase) (ETK) (NTK38).	Cytosolic phospholipase A2 (CPLA2) [Includ	2-acylhydrolase); Lysophospholipase (EC 3.1.1.5)]			DNA (cytosine-5)-methyltransferase 3A (EC	(DNA MTase HsalilA) (M.HsalilA).	DNA (cytosine-5)-methyltransferase 3B (EC	(DNA MTase HsalilB) (M.HsalilB).	DNA (cytosine-5)-methyltransferase-like prot	HsallP) (DNA MTase homolog HsallP) (M.Hs	DNA mismatch repair protein Mlh3 (MutL pro	DNA polymerase delta subunit 2 (EC 2.7.7.7).	DNA polymerase epsilon subunit B (EC 2.7.7.7) (DNA polymerase II subunit B).	UNA polymerase gamma subunit 2, mitochor polymerase accessory enhant? (DolG heta) //		Subarriit) (pos). DNA tonoisomerase II. alnha isozvme (FC 5 99 1.3)		DNA topolsomerase II, beta Isozyme (EC 5.99.1.3). Double-stranded RNA-hinding profein Staufen homolog		Dual specificity mitogen-activated protein kins	activating kinase 1) (c-Jun N- terminal kinase	Pual specificity protein phosphatase 1 (EC 3)	(MKP-1) (Protein-tyrosine phosphatase CL100	
51 CTA3 HIMANI	51 CIA3 HUMAN	NEW TOWAR	54 11385644	55 SRA4 RAT	56 CG1C_HUMAN	57 CCT1_HUMAN		58 CPAC_MOUSE		59 BMX_HUMAN		60 PA24 HUMAN	5NTC_HUMAN	62 1616601		63 DM3A_HUMAN		64 DM3B_HUMAN		65 DNM2_HUMAN				•	60 DDC2 HIMAN		_	72 STALL HIMAN			73 MPK4_HUMAN a		74 DUS1_HUMAN (

362 SOFV <u>FSFP</u> VSVGVHS 134 ALLN <u>FFFP</u> DEKPYSE	676 MSYG <u>FLFP</u> PYLSSSP 322 WYSE <u>YRFP</u> EELTQTF 167 GSHL <u>FGFP</u> PTPPKEV	237 REKD <u>YDFP</u> PPMRQAG 48 KPTV <u>FNYP</u> EGAAYEF 25 AKLG <u>FCFP</u> DLALQGD	370 PSTLEQEPTLLNGHM 394 ANTLEQEPSVLNSHG 138 GGSHEREPPSTPSEV 5 ADTGEAEPDWAYKPE	331 GKER <u>YNFP</u> NPPVE 331 GKER <u>YNFP</u> NPNPFVE	176 QPFH <u>EYFP</u> TPTVLCN 404 NNTG <u>FAFP</u> SDWCSNI 170 QQCT <u>FRFP</u> STAIKIQ 244 DSSR <u>FSYP</u> ERPIIFL 1451 LPRA <u>FAFP</u> VDPQVQS	209 TEGK <u>FSFP</u> TIHAIWS 350 NYTE <u>FKFP</u> QIKAHPW 88 DQGF <u>FLYP</u> KKISQAS 104 NLPN <u>FDFP</u> PEFYEHA	136 NVPD <u>FDFP</u> PEFYEHA 207 DEES <u>FQYP</u> SQQATVK 1054 VSYE <u>FKFP</u> FRNNNKW
		Enhancer of filmentation 1 (HE (PP105) (Neural precursor cell Estrogen receptor (ER) (Estrad Ets translocation variant 2 (Ets		フフ		- 85	Guanine nucleotide-binding protein G(S), alpha subunit (Adenylate cyclase-sumulating G alpha UMAN protein). JMAN High-mobility group protein 2-like 1 (HMGBCG protein). high-risk human papilloma viruses E6 oncoproteins targeted protein E6TP1 alpha; putative GAP high-risk human papilloma sapiens]
75 DUS4_HUMAN 76 EDA_HUMAN	77 NPP2_HUMAN 78 EF1G_HUMAN 79 GAT2_HUMAN	80 CASL_HUMAN 81 ESR1_MOUSE 82 ETV2_HUMAN	83 ELK3_HUMAN 84 ELK4_HUMAN 85 ERF_HUMAN 86 ERF_HUMAN	87 ETV3_HUMAN 88 ETV3_HUMAN 89 IF33_HUMAN 90 IF37_HUMAN	91 FOL1_HUMAN 92 FXJ2_HUMAN 93 FXK1_MOUSE 94 FZD4_HUMAN 95 GCP6_HUMAN	96 GGPP_HUMAN 97 KG3A_HUMAN 98 GDF3_HUMAN 99	100 GBAS_HUMAN 101 HM21_HUMAN 102 41513

194 KYGN <u>YFFP</u> GTGDMYE 133 PPKT <u>EAFP</u> YFEGSFQ 20 TGDT <u>EYFP</u> NFRASGA 97 GGPL <u>YPFP</u> RTVNDYT 1067 AQAP <u>YSFP</u> HNSPSHG 599 FPVR <u>FPYP</u> CTQTELA 3 ~ 52 RGAS <u>FLFP</u> PAESLSP	422 QGVV <u>FVFP</u> GGPGGLG 364 IYSS <u>FGFP</u> RTVKHID	46 NTRD <u>EMEP</u> GPNQMSG 134 STCS <u>ESYP</u> IRAGGDP 325 EPTK <u>EPEP</u> NKDSQLV 477 PLLTYREPPKFTLKA	334 SSSK <u>FNFP</u> SGRPGMQ 530 SVPD <u>FRFP</u> MADGHTD	76 VNRREHFPIYYLMAN 57 KNRK <u>EHFP</u> FYYLLAN -	367 SIHS <u>FGFP</u> NFVKKID	280 RMGQ <u>YEFP</u> NPEWSEV	140 RASL <u>YSFP</u> EPEAAQP 141 GNWQ <u>YFFP</u> VIFSKAS	168 MLSN <u>FGYP</u> GYQSKHY 191 PLNY <u>YNFP</u> KSCCTSV	37 KPCC <u>YIFP</u> GGRGDSA	113 KNTS <u>FAYP</u> AIRYLLY 259 MTGS <u>FEFP</u> EEEWSQI	315 YMSI <u>YSFP</u> MDEPISS 800 HRPL <u>FAFP</u> DAVKQIL
Histone deacetylase 3 (HD3) (RPD3-2). Homeobox protein DBX1. Homeobox protein Hox-C12 (Hox-3F). Homeobox protein Hox-C12 (Hox-3F). Homeobox protein PRH (Hematopoietically expressed homeobox) (Homeobox protein HEX). Homeodomain-interacting protein kinase 2 (EC 2.7.1). IkappaB kinase complex-associated protein (IKK complex-associated protein) (p150). Inositol-trisphosphate 3-kinase B (EC 2.7.1.127) (Inositol 1,4,5- trisphosphate 3-kinase) (IP3K) (IP3 3-kinase) (IP3K-B).	Integrin alpha-5 precursor (Fibronectin receptor alpha subunit) (Integrin alpha-F) (VLA-5) (CD49e). Interstitial collagenase precursor (EC 3.4.24.7) (Matrix metalloproteinase-1) (MMP-1) (Fibroblast collagenase).	72		Lysophosphatidic acid receptor Edg-2 (LPA receptor 1) (LPA-1). Lysophosphatidic acid receptor Edg-7 (LPA receptor 3) (LPA-3). Macrophage metalloelastase precursor (EC 3.4.24.65) (HME) (Matrix metalloproteinase-12) (MMP-	12) (Macrophage elastase) (ME). MAPK-activated protein kinase 2) (MAPKAP MAP kinase-activated protein kinase 2)	kinase 2) (MAPKAPK-2). Melanoma antigen preferentially expressed in tumors (Preferentially expressed antigen of	melanoma) (OPA-interacting protein 4) (OIP4). Melanoma-associated antigen 3 (MAGE-3 antigen) (Antigen MZ2-D). Metalloproteinase inhibitor 3 precursor (TIMP-3) (Tissue inhibitor of metalloproteinases-3) (MIG-5	protein). Methionine aminopeptidase 1 (EC 3.4.11.18) (MetAP 1) (Peptidase M 1) (Fragment). Microti ibi ile-associated protein 1A (MAP 1A) (Proliferation-related protein p80) [Contains: MAP1	light chain LC2]. Mitochondrial 28S ribosomal protein S29 (S29mt) (MRP-S29) (Death- associated protein 3) (DAP-3)		(MAP kinase isoform p97) (p97-MAPK). Mitogen-activated protein kinase kinase 6 (EC 2.7.1).
103 HDA3_HUMAN 104 DBX1_MOUSE 105 HXCC_HUMAN 106 HMPH_HUMAN 107 HIK2_HUMAN 108 IKAP_HUMAN	110 ITA5_HUMAN	112	116 432434 117 930341	119 EDG7_HUMAN	120 MM12_HUMAN	121 MKK2_HUMAN	122 MAPE_HUMAN 123 MAG3_HUMAN	124 TIM3_HUMAN 125 AMP1_HUMAN	126 MAPA_HUMAN	127 RT29_HUMAN 128 3133291	129 MK06_HUMAN 130 M3K6_HUMAN

131 TAB1_HUMAN	Mitogen-activated protein kinase kinase kinase 7 interacting protein 1 (TAK1-binding protein 1). Mitogen-activated protein kinase kinase kinase kinase 2 (EC 2.7.1.37)	361 LVRN <u>FGYP</u> LGEMSQP
132 M4K2_HUMAN 133 MDR1_HUMAN 134 MDR1_HUMAN	kinase 2) (MEK kinase kinase 2) (MEKKK 2) (Germinal center kinase) (GC kinase) (Rab8 interacting protein) (B lymphocyte serine/threonine protein kinase). Multidrug resistance protein 1 (P-glycoprotein 1) (CD243 antigen). Multidrug resistance protein 1 (P-glycoprotein 1) (CD243 antigen). Multidrug resistance protein 3 (P-alycoprotein 3)	737 PELT <u>FDFP</u> IETVVCL 1038 GEVV <u>ENYP</u> TRPDIPV 395 RNVH <u>FSYP</u> SRKEVKI 88 TAGNFSFPVNFSLSL
136 MDR3_HUMAN	Multidrug resistance protein 3 (P-glycoprotein 3)	397 NDVHESYPSRANVKI
137 DRNL_HUMAN 138 6959304	Muscle-specific DNase I-like precursor (EC 3.1.21) (DNase X) (XIB). MYB-binding protein 1A fHomo sapiens]	497 TKHPESFPLENGARE
139 2706549		135 SEDLFPFPMHGHSGG 751 RGAAFGFPGASPRAS
	Nyberii XV (Chronicaling in 1955) - 197. Neutrophil collagenase precursor (EC 3.4.24.34) (Matrix metalloproteinase-8) (MMP-8) (PMNL	
141 MM08_HUMAN	collagenase) (PMNL-CL). Nuclear factor NF-kappa-B p105 subunit (DNA-binding factor KBF1) (EBP- 1) [Contains: Nuclear	364 DISN <u>YGFP</u> SSVQAID
142 KBF1_HUMAN	factor NF-kappa-B p50 subunit]. Nuclear factor NF-kappa-B p105 subunit (DNA-binding factor KBF1) (EBP- 1) [Contains: Nuclear	405 SFPH <u>YGFP</u> TYGGITF
143 KBF1_HUMAN	factor NF-kappa-B p50 subunit].	400 TGPGYSFPHYGFPTY
144 RI14_HUMAN 145 RI14_HIMAN	Nuclear factor RIP140 (Nuclear receptor interacting protein 1). Nuclear factor RIP140 (Nuclear receptor interactino protein 1).	996 DNRIFSYPGVVKIPV 905 NDLEFKYPAGHGSAS
146 N153 HUMAN	Nuclear pore complex protein Nup153 (Nucleoporin Nup153) (153 kDa nucleoporin).	421 RESGESYPNFSLPAA
147 NCR1_HUMAN	Nuclear receptor co-repressor 1 (N-CoR1) (N-CoR).	23 HSVQYTFPNTRHQQE
148 RORG_HUMAN	Nuclear receptor ROR-gamma (Nuclear receptor RZR-gamma).	540 PPSPFSFPMNPGGWS
1_HUMAN	Nucleolar RNA helicase II (Nucleolar RNA helicase Gu) (RH II/Gu) (DEAD-box protein 21). Nucleoside diphosphate kinase homolog 5 (NDK-H 5) (NDP kinase homolog 5) (nm23-H5) (Testis-	136 RGVI <u>FLFP</u> IQAKIFH
150 NDK5_HUMAN	specific nm23 homolog) (Inhibitor of p53-induced apoptosis-beta) (IPIA-beta).	137 REIR <u>FMFP</u> EVIVEPI
151 OXE2_HUMAN	Olfactory receptor 51E2 (Prostate specific G-protein coupled receptor) (HPRAJ).	150 RGSLEFFPLPLLIKR
ORC4_HUMAN	Origin recognition complex subunit 4.	217 LMNSFGFPQYVKIFK
SE_HUMAN		24 ERHHFSFPSIFIYGH
154 2645205	p160 myb-binding protein [Mus musculus] Dodilia	493 INCHESTEDDRING 414 FEHVYSFDNKOKSAF
	Faxiiii. Peptidyl-prolyl cis-trans isomerase like 2 (EC 5.2.1.8) (PPIase) (Rotamase) (Cyclophilin-60)	
156 CYP6_HUMAN	(Cyclophilin-like protein Cyp-60). Perinheral nlasma memhrane protein CASK (FC 2 7 1 -) (hCASK) (Calcium/calmodulin-dependent	45 SLQP <u>FVYP</u> VCTPDGI
157 CSKP_HUMAN	serine protein kinase) (Lin-2 homolog).	763 HPDR <u>FAYP</u> IPHTTRP
	Peroxisomal 3,2-trans-enoyl-CoA isomerase (EC 5.3.3.8) (Dodecenoyl-CoA delta-isomerase) (D3,D2-enoyl-CoA isomerase) (DBI-related protein 1) (DRS-1) (Hepatocellular carcinoma-	
158 PECI_HUMAN	associated antigen 88).	246 GCSS <u>YTFP</u> KIMSPAK

417 PDDI <u>FLFP</u> KLLQKMA	389 PDSQ <u>YLFP</u> KLLQKMA	269 SPVLFRFPAASSDNT	628 GLDL <u>FVFP</u> YRVVATA	237 KFMY <u>FEFF</u> QPLPVCG	608 ELLD <u>enyp</u> dqyvrey	581 ELLDESFPDCHVGSF	161 AWL <u>WYSFP</u> LULEFSA 366 ADEY <u>FTFP</u> KGPVDET	580 ELLDESFPDCYVGSF	213 PDAFYYFPDSGFFCE	376 TGPAFTFPSDVPYQA	176 YVGR <u>FKFP</u> EERAAEV	62 GYVN <u>FRFP</u> ADAEWAL	402 SELE <u>FOFF</u> DUKDEVW	587 KGSAFTFPSQQSPRN	601 KKGEYVFPAKKKLQE	552 KNVC <u>FIFP</u> QFLYQFF	485 VYNSFGFFGGAARIF	354 VFEIEWEFCGNOERN 405 VAINEDEPKI AETYL	186 VFVGFLFPWALMLLS	400 ELVGELEDICITIAS	446 AFITEMEPEHAVKAY	455 CEARTULINGSAGVUR	1805 ENDVEREPREI DMFP	
Peroxisome proliferator activated receptor alpha (PPAR-alpha).	Peroxisome proliferator activated receptor delta (PPAR-delta) (PPAR-beta) (Nuclear hormone recentor 1) (NUC1).	i 3-kinase Pi3K).	Phosphatidylinositol 4-kinase alpha (EC 2.7.1.67) (PI4-kinase) (PtdIns-4-kinase) (PI4K-alpha). Phosphatidylinositol-3.4,5-trisphosphate 3-phosphatase PTEN (EC 3.1.3.67) (Mutated in multiple	advanced cancers 1). Phosnhatidylinositol 4.5-bisphosphate 3-kinase catalytic subunit, beta isoform (EC 2.7.1.153) (PI3-	kinase p110 subunit beta) (PtdIns-3-kinase p110) (PI3K) (PI3Kbeta). Phosphatidylinositol-4 5-bisphosphate 3-kinase catalytic subunit, delta isoform (EC 2.7.1.153) (PI3-	kinase p110 subunit delta) (PtdIns- 3-kinase p110) (PI3K) (p110delta). Phosphatidylinositol-4,5-bisphosphate 3-kinase catalytic subunit, delta isoform (EC 2.7.1.153) (PI3-		22758919 phosphoinositide 3 kinase P110delta [Mus musculus]	2696236 phospholipase B [Rattus norvegicus]		is Plenty of Shost, POSH [muscalus] Polyadenviate-binding protein 5 (Poly(A)-binding protein 5) (PABP 5).	Polyadenylate-binding protein 5 (Poly(A)-binding protein 5) (PABP 5).		Polyposis locus protein 1 (TB2 protein).	Potassium voltage-gated channel subramily NQT IIIEIIIDELS (FOtassium channel NQT-IIIC S):	Potential melicase in CVT of Potential Properting ATPase IS (EC 3.6.3.1) (Fragment).		binding protein) (MTB-ZF).	Probable ATP-dependent KNA helicase p54 (Oncogene KOA) (DEAD-50X protein 9).	Probable G protein-coupled receptor GPR68 (Ovarian cancer G protein- coupled receptor 1) (OGR-	1). Probable RNA-binding protein KIAA0682.	Probable tumor suppressor protein MN1.	(Ubiquitin-specific processing protease FAF-X) (Deubiquitinating enzyme FAF-X) (Fat facets protein	
159 PPAR HUMAN	160 PDAS MOLISE	164 D85A MOLISE	162 PI4K_HUMAN	163 PTEN_HUMAN	164 P11B_HUMAN	165 P11D_HUMAN	166 P11D_HUMAN		169 269623	PLEK_H	171 3002588 173 BABE HIMAN	173 PAB5 HUMAN	174 15667468	175 DP1_HUMAN	176 CIQ3_HUMAN	177 MV10_HUMAN 178 A11A_HUMAN	4097902	180 PRD2_HUMAN	181 DDX6_HUMAN	182 GPR4_HUMAIN	183 GP68_HUMAN 184 K682_HUMAN	185 MN1_HUMAN		

1616 RDDVEGYPHQFEDKP 183 PDHNELFPEFEIVIE 742 TDTLFVFPSREDATP 794 QNAA <u>FMYP</u> PNPWKEI 787 QNAA <u>FMYP</u> PNPWKEI 97 DQKS <u>FIFP</u> QESEGTF	234 DVFT <u>EGFP</u> VPPFLLE 262 RDRR <u>FHFP</u> EETPETP 261 RDRP <u>FHFP</u> EETPETP 172 TNNS <u>FAFP</u> ESNETQA	1704 ICTM <u>FYYP</u> QKIPNKP	247 AIGV <u>ELFP</u> AFLIASA 172 TNNT <u>FAFP</u> ESNETQA	1565 TSTD <u>FSFP</u> DVNEKDA	720 HYST <u>EAYP</u> PTEVTSH 4222 LYGG <u>FPFP</u> LEMENKR 100 NNQL <u>FRFP</u> ATSPLKT	244 NVAS <u>FLYP</u> NLGGSWR 251 IIPG <u>FPYP</u> TAATTAA 370 GNTP <u>FIFP</u> LYGHGEI	369 GNTP <u>ELFP</u> LYGQGEL	371 GNTP <u>FLFP</u> LYGQGEI 311 HKCA <u>FQFP</u> GSPPGGG 786 PRSP <u>YKFP</u> SSPLRIP 100 CDQR <u>FRFP</u> SPILKVQ	467 KENA <u>FPFP</u> MDNQFSM
Probable ubiquitin carboxyl-terminal hydrolase FAF-Y (EC 3.1.2.15) (Ubiquitin thiolesterase FAF-Y) (Ubiquitin-specific processing protease FAF-Y) (Deubiquitinating enzyme FAF-Y) (Fat facets protein 187 FAFY_HUMAN related, Y-linked) (Ubiquitin-specific protease 9, Y chro 188 PDC2_HUMAN Programmed cell death protein 2 (Zinc finger protein Rp-8). 189 3858885 proliferation potential-related protein [Mus musculus] 190 KPCM_HUMAN Protein kinase C, mu type (EC 2.7.1) (nPKC-mu) (Protein kinase EPK2). 191 KPCN_HUMAN Protein kinase C, nu type (EC 2.7.1) (nPKC-nu) (Protein kinase EPK2).	Protein phosphatase 1, regulatory subunit 3D (Protein phosphatase 1, regulatory subunit o) (Protein 193 PP3D_HUMAN phosphatase 1 binding subunit R6). 194 10567793 protein tyrosine phosphatase BK [Mus musculus] 195 1144002 protein tyrosine phosphatase D30 [Rattus norvegicus] 196 23268287 protein tyrosine phosphatase, non-receptor type 13 (EC 3.1.3.48) (Protein-tyrosine phosphatase 1E)	_		Protein-tyrosine pnospnatase zeta precursor (EC 3.1.3.40) (N-F 1F- zeta) (F103pnacan) (3F 3.00 PTPZ_RAT chondroitin sulfate proteoglycan) (3H1 keratan sulfate proteoglycan). Protein-tyrosine phosphatase zeta precursor (EC 3.1.3.48) (R-PTP- zeta) (Phosphacan) (3F8	201 PTPZ_RAT chondroitin sulfate proteoglycan) (3H1 keratan sulfate proteoglycan). 202 FAT2_HUMAN Protocadherin Fat 2 precursor (hFat2) (Multiple epidermal growth factor-like domains 1). 203 8216989 putative cell cycle control protein [Homo sapiens]	GP40_HUMAN RBM9_HUMAN 6007826	Rab proteins geranyigeranyiransierase component A 1 (Nab escort protein 1) (Nat - 1) 207 RAE1_HUMAN (Choroideraemia protein) (TCD protein). Rab proteins geranyideranyitransferase component A 2 (Rab escort protein 2) (REP-2)	208 RAE2_HUMAN (Choroideraemia-like protein). 209 RBPL_HUMAN Recombining binding protein suppressor of hairless-like protein (Transcription factor RBP-L). 210 RB_HUMAN Retinoblastoma-associated protein (PP110) (P105-RB) (RB). 211 RBB5_HUMAN Retinoblastoma-binding protein 5 (RBBP-5) (Retinoblastoma-binding protein (RBBP-8) (CtBP interacting protein (CtIP) (Retinoblastoma-binding protein 8 (RBBP-8) (CtBP interacting protein) (CtIP) (Retinoblastoma-binding protein 8 (RBBP-8) (CtBP interacting protein) (CtIP) (Retinoblastoma-binding protein 8 (RBBP-8) (CtBP interacting protein)	212 RBB8_HUMAN interacting protein and myosin-like) (RIM).

170 FQDI <u>FKYP</u> QEEQPRQ 1830 PISL <u>ESFP</u> PLLPQQF 505 KDQK <u>YIFP</u> TLDKPSV	34 LYED <u>FLFP</u> IDISLVK 163 PPPA <u>FTYP</u> ASLHAQM	450 SSGS <u>YQFP</u> MVPGGDR 66 AASL <u>FGFP</u> FQLTTKP 437 VFST <u>FFYP</u> KLKSGGY 195 EIHI <u>YQFP</u> ECDSDED	314 GIKI <u>YQFP</u> DCDSDED 214 GIHV <u>YQFP</u> ECDSDED 200 KIKI <u>YEFP</u> ETDDEEE	342 SRVE <u>FTFP</u> DFVTEGA 225 RQVD <u>FKFP</u> SSVPAGA 371 EFRT <u>YSFP</u> CYLPQPL	708 NIPR <u>EYFP</u> EGLPDTC 96 SLHS <u>YPFP</u> GTIKSRD 432 KFSPFRFPDSGLPVS	232 SSMN <u>FRFP</u> QCVPINL 421 EFKN <u>YNFP</u> KYKPEPL 627 AVPS <u>FDFP</u> KTPSSQN 664 SYLI <u>YVFP</u> DRPKDEV 664 NYLI <u>YVFP</u> DRPKDEV	333 REGF <u>YLFP</u> DGRNQNP 966 ILRE <u>FAFP</u> PVSPRL 2225 SSSS <u>FPFP</u> CKAWPSG 1162 SFRS <u>YYFP</u> VKNVIDG
 IUMAN Retinoblastoma-like protein 2 (130 kDa retinoblastoma-associated protein) (PRB2) (P130) (RBR-2). 12053793 retinoid-acid induced protein 1 [Homo sapiens] Rho guanine nucleotide exchange factor 2 (GEF-H1 protein) (Proliferating cell nucleolar antigen P40). Rho-GTPase-activating protein 7 (Rho-type GTPase-activating protein 7) (Deleted in liver cancer 1 protein) (Dic-1) (HP protein) (StAR-related lipid transfer protein 12) (StARD12) (START domain- 				Serine/threonine kinase b (EC (Aurora-related kinase 1) (hAF Serine/threonine protein kinas Serine/threonine protein kinas Serine/threonine protein phosi		kinase RCK). Serine/threonine-protein kinase MAK (EC 2.7.1) (Male germ cell- associated kinase). N Serine/threonine-protein kinase PCTAIRE-2 (EC 2.7.1). N Serine/threonine-protein kinase ULK1 (EC 2.7.1) (Unc-51-like kinase 1). N Signal transducer and activator of transcription 5A.	54
213 RBL2_HUMAN 214 120537 215 ARH2_HUMAN	216 RHG7_HUMAN 217 RBMS_HUMAN	218 RUNZ_HUMAN 219 SENZ_HUMAN 220 SENZ_HUMAN 221 SEP1_HUMAN	222 SEP4_HUMAN 223 SEP5_HUMAN 224 SEP7_HUMAN	225 STK6_HUMAN 226 STKD_MOUSE 227 KPT3_HUMAN	228 2ACA_HUMAN 229 ST19_HUMAN	230 MAK_MOUSE 231 MAK_HUMAN 232 KPT2_HUMAN 233 ULK1_HUMAN 234 ST5A_HUMAN	236 CBL_HUMAN 237 CBLB_HUMAN 238 6649242 239 S3B3_HUMAN

WO 2005/007090		PCT/US2004/021514
253 SNSVETYPENGTDDF 186 GVKAESEPETVFTTV 205 EFRTEIFPETVFTAV 597 FGSLEPYPYTYMAAA 235 GMASEREPETTFISV 106 SSANFTFPGYPIHVP 65 QESRELYPGKNGRLG 270 NAKEFIEPNMQGQGS 147 SPHLETFPTPPKDV 104 VSPRESFPGTTGSLA 365 EPEDEAFPSTAPSPQ 345 APQPYTFPASLSTIN	203 TALQETYPLFTTNAC 729 PPENYDEPVVIVKQE 52 GGAAFIEPNTSVYPE 767 GFGSFRFPSGNQGGA 220 GLRRFAFPLSLFQGS 120 LDASFRYPQDYQFYI 2437 RPGSFTFPGDSDSLQ 947 SKDKFEFPLTPVGEE	114 YRIR <u>FYFP</u> RWYCSGS 100 YRIR <u>FYFP</u> NWFGLEK 292 RIKS <u>YSFP</u> KPGHRKS 785 RYPN <u>YMFP</u> SQGITPQ 230 SFCH <u>FVFP</u> LCDARSR 223 SKPG <u>FGFP</u> FETNYPE 330 ISLD <u>FTYP</u> SLEESIP 537 DSD <u>YFKY</u> PLMALGEL
Stac protein (SRC homology 3 and cysteine-rich domain protein). T-box transcription factor TBX18 (T-box protein 18) (Fragment). T-box transcription factor TBX20 (T-box protein 20) (Fragment). T-box transcription factor TBX3 (T-box protein 3). T-box transcription factor TBX6 (T-box protein 6). TFIIA-alpha and beta like factor (ALF). TFIIA basal transcription factor complex p34 subunit (Basic transcription factor 2 34 kDa subunit) (BTF2-p34) (General transcription factor IIH polypeptide 3). Tob1 protein (Transducer of erbB-2 1). Trans-acting T-cell specific transcription factor GATA-3. Transcription factor GATA-4 (GATA binding factor-4). Transcription factor GATA-5 (GATA binding factor-5). Transcription factor p65 (Nuclear factor NF-kappa-B p65 subunit).		Tyrosine-protein kinase JAK2 (EC 2.7.1.112) (Janus kinase 2) (JAK-2). Tyrosine-protein kinase JAK3 (EC 2.7.1.112) (Janus kinase 3) (JAK-3) (Leukocyte janus kinase) (L-JAK). Tyrosine-protein kinase SYK (EC 2.7.1.112) (Spleen tyrosine kinase). Tyrosine-protein kinase transmembrane receptor ROR1 precursor (EC 2.7.1.112) (Neurotrophic tyrosine kinase, receptor-related 1). Tyrosine-protein kinase transmembrane receptor ROR2 precursor (EC 2.7.1.112) (Neurotrophic tyrosine kinase, receptor-related 2). Ubiquitin carboxyl-terminal hydrolase 26 (EC 3.1.2.15) (Ubiquitin thiolesterase 26) (Ubiquitin-specific processing protease 26) (Deubiquitinating enzyme 26). Ubiquitin carboxyl-terminal hydrolase 8 (EC 3.1.2.15) (Ubiquitin thiolesterase 8) (Ubiquitin-specific processing protease 8) (Deubiquitin-fusion degradation protein 2).
240 STAC_HUMAN 241 TX18_HUMAN 243 TX20_HUMAN 244 TBX6_HUMAN 245 T2AY_HUMAN 246 TFH3_HUMAN 247 TOB1_HUMAN 248 GAT3_HUMAN 249 GAT4_HUMAN 250 GAT5_HUMAN	252 TCL5_HUMAN 253 TF1A_HUMAN 254 ERG_MOUSE 255 TERA_HUMAN 256 E2BE_HUMAN 257 SSRA_HUMAN 258 TRIO_HUMAN 259 1113923	262 JAK2_HUMAN 263 KSYK_HUMAN 264 ROR1_HUMAN 265 ROR2_HUMAN 266 UBPQ_MOUSE 267 UBPR_HUMAN 268 UBPR_HUMAN

80 EDPA <u>FGFP</u> KLEQANK	274 DISY <u>EGYP</u> SFRRSSL 1001 GFQA <u>YDFP</u> AVTTAQY	921 YKEL <u>FSYP</u> KHITSNT 565 RRTG <u>FSFP</u> TQEPRPQ 352 GRRD <u>FVYP</u> SSTRDPS 577 KSLC <u>FQYP</u> PVYVGKI	270 PPAE <u>FSYP</u> VDNQRGS	218 KLGP <u>FGYP</u> PTLVYQN 149 PSGA <u>YVFP</u> PPVANGM	342 CGKG <u>FDFP</u> GSARIHE 792 LNPT <u>FTFP</u> SHSLTQS 44 NFRN <u>FPYP</u> DLAGPRK	A series	Acid Target Sequence	395 PPSD <u>FLFP</u> RPNRFQP 679 ITSL <u>FFF</u> TSSDCPF 167 GSHL <u>FGFP</u> PTPPKEV 330 HAQP <u>FDFP</u> DDNQNSK 136 NVPDFDFPPEFYEHA	599 FPVR <u>FPYP</u> CTQTELA 751 GLETYGFPSVTLISC 306 STNP <u>FDFP</u> FVSQGEV	80 FPQGYQFPSMEQLAA 358 PMDRFSFPDGKESDT 86 DVADYVFPAQGDNSF 89 DTADYTFPLQGNSFF 186 VFVGFLFPWALMILS
269 27434480 ubiquitin ligase E3 alpha-II [Homo sapiens]	Upiquitinprotein ligase EDD (EC 0.3.2) (hiyperplastic discorption from 270 EDD_HUMAN induced protein). 271 SYV2_HUMAN Valyf-tRNA synthetase 2 (EC 6.1.1.9) (ValinetRNA ligase 2) (ValRS 2) (G7a).	272 PPOV_HUMAN related/lalphal-related H5/proline-rich) (PH5P). 273 VINE_HUMAN Vinexin (SH3-containing adaptor molecule-1) (SCAM-1). 274 VINE_HUMAN Vinexin (SH3-containing adaptor molecule-1) (SCAM-1). 275 WRN_HUMAN Werner syndrome helicase.	7	277 WAS2_HUMAN homology domain-containing protein 2). 278 WBP2_MOUSE WW domain binding protein 2 (WBP-2). 7inc finger protein 44 (Zinc finger protein KOX7) (Gonadotropin inducible transcription repressor-2)	279 ZN44_HUMAN (GIOT-2). 280 HRX_HUMAN Zinc finger protein HRX (ALL-1) (Trithorax-like protein). 281 Z287_HUMAN Zinc finger protein ZNF287.	TABLE 3: CARDIOVASCULAR	Accession Code		5 GBAS_HUMAN protein). 6 IKAP_HUMAN lkappaB kinase complex-associated protein (IKK complex-associated protein) (p150). 7 LGR6_HUMAN Leucine-rich repeat-containing G protein-coupled receptor 6. 8 MYHD_HUMAN Myosin heavy chain, skeletal muscle, extraocular (MyHC-eo).	9 NDR4_HUMAN protein-8) (SMAP-8). 10 ACHO_RAT Neuronal acetylcholine receptor protein, beta-3 chain precursor. 11 P2X1_HUMAN P2X purinoceptor 1 (ATP receptor) (P2X1) (Purinergic receptor). 12 P2X7_HUMAN P2X purinoceptor 7 (ATP receptor) (P2X7) (Purinergic receptor) (P2Z receptor). 13 GPR4_HUMAN Probable G protein-coupled receptor GPR4 (GPR19).

190 FLVG <u>FLEP</u> ICLLLAS 225 LAVA <u>ETEP</u> FITTVTC 244 NVAS <u>FLYP</u> NLGGSWR 244 APYH <u>ESFP</u> LDFLVKS 734 KPDL <u>FTYP</u> CEMSLIS 200 NILG <u>FLFP</u> FLIILTS 435 FGTE <u>VVFP</u> EVLLTEI 1971 PYGV <u>FIFP</u> NKTRPLS 879 DGLK <u>YLFP</u> GECQYVL 431 FSPR <u>FPF</u> TVPPAPG	Amino Target Sequence Acid	116 ELMDEGFPOTTDSKI 97 SGETESYPDFLRMML 23 QVVTETEPFGFQGIS 947 FYCLYSFPSKKSKAR 102 GGGDFDYPGAPAGPG 84 DPRPFAYPSHTFGPD 915 ARGSFEFPVGDAVSK 201 THCQYNFPQVGRTAL 167 GSHLFGFPTPPKEV 125 SKPTFPWPPGNKPSL 88 KKDKFAFPVPYGLGS 599 FPVRFPYPCTQTELA 288 DQVLFPYPEDNGQRK 134 RVCTFRFPSTNIKIT 280 QFHSFIYPYMANGSL 249 MMPPFMYPPYLPFPP
Probable G protein-coupled receptor GPR68 (Ovarian cancer G protein- coupled receptor 1) (OGR- 1). HUMAN Putative G protein-coupled receptor GPR40. Putative P2Y purinoceptor FKSG79. HUMAN Relaxin receptor 1 (Leucine-rich repeat-containing G protein-coupled receptor 7). Type-1 angiotensin II receptor (AT1) (AT1AR). Vascular non-inflammatory molecule 2 precursor (Vanin 2) (Glycosylphosphatidyl inositol-anchored protein GPI-80) (FOAP-4 protein). 16904210 very large G protein-coupled receptor 1 [Mus musculus] Von Williebrand factor precursor (WWF). Williams-Beuren syndrome chromosome region 14 protein (WS basic-helix- loop-helix leucine zipper protein) (WS-bHLH) (Mix interactor).	TABLE 4: INFLAMMATION de Target Description	Adaptor-related protein complex 1, mu 2 subunit (Mu-adaptin 2) (Adaptor protein complex AP-1 mu-2 subunit) (Golgi adaptor HA1AP1 adaptin mu-2 subunit) (Clathrin assembly protein complex 1 medium chain 2) (AP-mu chain family member mu1B). Allograft inflammatory factor-1 (AIF-1) (Ionized calcium-binding adapter molecule 1) (G1). Apolipoprotein L3 (Apolipoprotein L-III) (ApoL-III) (TNF-inducible protein CG12-1) (CG12_1). Calcineurin-binding protein Cabin 1 (Calcineurin inhibitor) (CAIN). CCAAT/enhancer binding protein alpha (C/EBP alpha). CCAAT/enhancer binding protein alpha (C/EBP alpha). CCAAT/enhancer binding protein alpha (C/EBP alpha). CCAAT/enhancer binding protein alpha (C/EBP applan). C-X-C chemokine receptor type 3 (CXCR-3) (CXCR-3) (CKR-L2) (CD183 antigen). Endothelial transcription factor GATA-2. FYN-binding protein (FYN-T-binding protein) (FYB-120/130) (p120/p130) (SLP-76 associated phosphoprotein) (SLAP-130). 70 HMG-box protein SOX21 [Mus musculus] IkappaB kinase complex-associated protein (IKK complex-associated protein) (p150). Interferon regulatory factor 4 (IRF-4) (Lymphocyte specific interferon regulatory factor) (LSIRF) (NF-EM5) (Multiple myeloma oncogene 1). Interleukin enhancer-binding factor 1 (Cellular transcription factor ILF-1). Interleukin-1 receptor-associated kinase-2 (EC 2.7.1) (IRAK-2). Large proline-rich protein BAT2 (HLA-B-associated transcript 2) (G2).
14 GP68_HUMAN 15 GP17_HUMAN 16 GP40_HUMAN 17 FK79_HUMAN 18 LGR7_HUMAN 19 AG2R_HUMAN 20 VNN2_HUMAN 21 1690421 22 VWF_HUMAN 23 WS14_HUMAN	Accession Code	1 A1M2_HUMAN 2 AIF1_HUMAN 3 APL3_HUMAN 4 CABI_HUMAN 5 CEBA_HUMAN 6 CEBE_RAT 7 CO4_HUMAN 8 CCR3_HUMAN 9 GAT2_HUMAN 11 Z8274770 12 IKAP_HUMAN 13 IRF4_HUMAN 14 ILF1_HUMAN 15 IRA2_HUMAN 16 BAT2_HUMAN

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565 RRTG <u>FSFP</u> TQEPRPQ 352 GRRD <u>FVYP</u> SSTRDPS 1255 KSEK <u>FSWP</u> QRSETLS	Amino Target Sequence Acid Target Sequence 777 DEEPEDFYVLPTL 273 DLPKYLFPEDPSYSS 271 DLPSYLFPEDPSYDA	84 TDQVEPYPSVLNEEQ 359 SRDYYNEPLALAGPP 547 LPIDYYEPPQKTCLI 4198 NFPREQEPGKPGIYT 23 QVVTETEPFGFQGIS 31 SALGEKYPVGNNQTA 78 NNPCERYPTPGEAPG 379 KTLSFYFEPCGKIPP 212 PYESETTEPELMRKGS 1081 VDCKETYPSRPDSQV 158 EEERFGFPAFSGISR 27 SSAAFGFPRGAGPSQ 27 SSAAFGFPRGAGPSQ 27 SSAAFGFPRGAGPSQ 27 SSAAFGFPRGAGPSQ 27 SSAAFGFPRGAGPSQ 28 DPRPFAYPPHTFGPD 1339 QRKEFNFPEAGSSSG 286 NIFSYLFPKYSTNEA 280 EEVSYYFPLKTLWRS 223 RKIQENFEPSEMTGIW 442 ETLSFIFPEGIVAGG 638 KYRDFEEPSEMTGIW 474 LQMDFGFPEHLLVDF	
Vinexin (SH3-containing adaptor molecule-1) (SCAM-1). Vinexin (SH3-containing adaptor molecule-1) (SCAM-1). Zinc finger protein 40 (Human immunodeficiency virus type I enhancer- binding protein 1) (HIV-EP1) (Major histocompatibility complex binding protein 1) (MBP-1) (Positive regulatory domain II binding factor 1) (PRDII-BF1).	TABLE 5: METABOLIC DISORDERS 1-phosphatidylinositol-4,5-bisphosphate phosphodiesterase beta 3 (EC 3.1.4.11) (Phosphoinositide phospholipase C) (PLC-beta-3) (Phospholipase C-beta-3). 5'-AMP-activated protein kinase, catalytic alpha-1 chain (EC 2.7.1) (AMPK alpha-1 chain). 5'-AMP-activated protein kinase, catalytic alpha-2 chain (EC 2.7.1) (AMPK alpha-2 chain).	Acyl-CoA dehydrogenase, very-long-chain specific, mitochondrial precursor (EC 1.3.99-) (VLCAD). Androgen receptor (Dihydrotestosterone receptor). Androgen receptor (Dihydrotestosterone receptor). Androgen receptor (Dihydrotestosterone receptor). Apolipoprotein B-100 precursor (Apo B-100) (Contains: Apolipoprotein B-48 (Apo B-48)]. Apolipoprotein L3 (Apolipoprotein L-III) (Apol-III) (TNF-inducible protein CG12-1) (CG12_1). Aquaporin-CHIP (Water channel). ATP-binding cassette, sub-family, A, member 1 (ATP-binding cassette transporter 1) (ATP-binding cassette); Sub-family, A, member 2 (Iron inhibited ABC transporter 1) (ATP-binding cassette) (Cholesterol efflux regulatory protein). ATP-binding cassette, sub-family F, member 2 (Iron inhibited ABC transporter 2) (HUSSY-18). Beta-hexosaminidase alpha chain precursor (EC 3.2.1.52) (N-acetyl- beta-glucosaminidase) (Beta-N-acetylhexosaminidase) (Hexosaminidase A). Bile salt export pump (ATP-binding cassette, sub-family B, member 11). Biliverdin reductase A precursor (EC 1.3.1.24) (Biliverdin-IX alpha- reductase). CCAAT/enhancer binding protein alpha (C/EBP alpha). CCAAT/enhancer binding protein alpha (C/EBP alpha). CCAAT/enhancer binding protein alpha (C/EBP epsilon). Cchat/fenhancer binding protein alpha (C/EBP epsilon). Chloride channel protein 3 (ClC-3). Chloride channel protein 6 (ClC-6). Chloride channel protein (C/C-CA). Chloride channel protein (C/C-CA). Chloride channel protein CLC-KB (ClC-K2). Chloride intracellular channel 6.	
46 VINE_HUMAN 47 VINE_HUMAN 48 ZEP1_HUMAN	Accession Code 1 PIP3_HUMAN 2 AAK1_HUMAN 3 AAK2_HUMAN	4 ACDV_HUMAN 5 ANDR_HUMAN 7 APB_HUMAN 8 APL3_HUMAN 10 ABC1_HUMAN 11 ABF2_HUMAN 12 HEXA_HUMAN 13 AB11_HUMAN 15 CEBA_HUMAN 16 CEBA_HUMAN 17 CEBE_HUMAN 17 CEBE_HUMAN 18 13562153 19 CLC3_HUMAN 20 CLC3_HUMAN 21 CLC6_HUMAN 21 CLC6_HUMAN 22 CICL_HUMAN 23 CLIC_HUMAN 23 CLIC_HUMAN 24 CETE_HUMAN	MANDEL TIES 47

W U 2005/00/0	170						PCT/U	S2004/0	21514	
163 LGFS <u>FSFP</u> CHQTGLD 598 LGFT <u>FSFP</u> CQQNSLD 599 FPVR <u>FPYP</u> CTQTELA 280 QFHS <u>FIYP</u> YMANGSL 78 VVSA <u>FGFP</u> VILARVA 142 MEEE <u>FNYP</u> LDNVHLL	1025 QCGL <u>FSFP</u> CKNGRCV 173 YPQQ <u>FPYP</u> QAPFVSQ 281 DDLL <u>YSFP</u> VVIKNKT 227 VRNT <u>YIFP</u> PEPSMKI	113 KNTS <u>FAYP</u> AIRYLLY 2 HGQT <u>ETFP</u> DLFPEKD 30 IGFS <u>YAFP</u> KSITVFF 30 IGFS <u>YAFP</u> KAVTVFF	29 IGFA <u>TGFF</u> KAVSVFF 32 TGFS <u>YAFP</u> KAVSVFF 306 STNP <u>FDFP</u> FVSQGEV	37 MVSD <u>FFYP</u> NMGGVES	24 FFFD GET ENVENEEN	75 HPQP <u>YIFP</u> DSPGG1S 452 GNNS <u>YVFP</u> GVALGVV 669 TFQF <u>YRFP</u> PATTPRL 1295 NAQOFPFPPNYGISQ	23 HSVQ <u>YTFP</u> NTRHQQE	2160 PAPL <u>YSFP</u> GASCPVL 540 PPSP <u>FSFP</u> MNPGGWS 365 QLLVFMFPVGLYYCF	513 CLNE <u>FNFP</u> DPYSKV <u>K</u> 417 PDDI <u>FLFP</u> KLLQKMA	389 PDSQ <u>YLFP</u> KLLQKMA
 Hexokinase type III (EC 2.7.1.1) (HK III). Hexokinase, type I (EC 2.7.1.1) (HK I) (Brain form hexokinase). Hexokinase, type II (EC 2.7.1.1) (HK II) (Muscle form hexokinase). IkappaB kinase complex-associated protein (IKK complex-associated protein) (p150). Interleukin-1 receptor-associated kinase-2 (EC 2.7.1) (IRAK-2). Leptin receptor gene-related protein (OB-R gene related protein) (OB- RGRP). Lipoprotein lipase precursor (EC 3.1.1.34) (LPL). 		Mitochondrial 28S ribosomal p (Ionizing radiation resistance c 75 mitochondrial ribosomal proteii Monocarboxylate transporter 1 Monocarboxylate transporter 2					Nuclear receptor coactivator z Nuclear receptor co-repressor Nuclear receptor co-repressor receptor) (SMRT) (SMRTe) (TI			
48 HXK3_HUMAN 49 HXK1_HUMAN 50 HXK2_HUMAN 51 IKAP_HUMAN 52 IRA2_HUMAN 53 OBRG_HUMAN 54 LIPL_HUMAN	55 LRP2_HUMAN 56 LOL1_HUMAN 57 MDHC_HUMAN 58 MUTA_HUMAN	59 RT29_HUMAN 60 141415i 61 MOT1_HUMAN 62 MOT2_HUMAN	63 MOT3_HUMAN 64 MOT4_HUMAN 65 MYHD_HUMAN	66 PIGA_HUMAN	67 NB8M_HUMAN	68 NIZM_HUMAN 69 MAOX_HUMAN 70 NPH4_HUMAN	71 NCO2_HUMAN 72 NCR1_HUMAN	73 NCR2_HUMAN 74 RORG_HUMAN	75 PNK4_HUMAN 77 PPAR_HUMAN	78 PPAS_MOUSE

581 ELLDESFPDCHVGSF 190 FSKA <u>FFFP</u> SFNVRDL 366 ADEY <u>FTFP</u> KGPVDET	140 IQGF <u>FSFP</u> VDNLRAS 75 TPCL <u>YKFP</u> DHTLSHG 3321 DKNK <u>FYFP</u> SLQPRKD 2512 NLVA <u>FPFP</u> HAAILED 2370 RYGL <u>FVYP</u> KFQPPWD 2687 PGLL <u>FHFP</u> RRSQKDC	1880 GYAL <u>YFFP</u> EQQRFNS 3060 SHVR <u>FVFP</u> EPTADVN	348 KATP <u>YTFP</u> GGTGQII 56 NLIG <u>FGYP</u> AYISIKA	153 VWLL <u>FEYP</u> ESSGPAR	236 VWLI <u>FEYP</u> ESSGSAR	156 KFFG <u>FKFP</u> GLRVLTY 942 ETNV <u>FFYP</u> RLLPLTK 370 GNTP <u>FIFP</u> LYGHGEI	43 GTSA <u>YAFP</u> SLGPVAL	22 PPYA <u>FFFP</u> MLGGLS 23 AGFP <u>FAFP</u> GALRGSP	370 PSQP <u>YMFP</u> RMLMKIT 1830 PISL <u>FSFP</u> PLLPQQF	39 VPCS <u>FSYP</u> WRSWYSS 120 GGGA <u>FMFP</u> YFIMLIF	279 IGPL <u>FFFP</u> LLYMIFQ
Phosphatidylinositol-4,5-bisphosphate 3-kinase catalytic subunit, delta isoform (EC 2.7.1.153) (Pl3-kinase p110 subunit delta) (Ptdlns- 3-kinase p110) (Pl3K) (p110delta). Phosphatidylinositol-glycan biosynthesis, class O protein (PIG-O). 1 phosphodiesterase A' subunit [Homo sapiens]	Phosphoribosyl pyrophosphate protein 1) (39 kDa phosphorib Pituitary-specific positive trans Polycystic kidney and hepatic Polycystic kidney and hepatic Polycystic kidney and hepatic Polycystic kidney and hepatic Polycystic kidney disease 1-lik		acetylgalactosaminyltransferase) (UDP-GalNAc:polypeptide, N- acetylgalactosaminyltransferase) (GalNAc-T1). Polyposis locus protein 1 (TB2 protein).	Potassium voltage-gated channel subfamily A member 1 (Potassium channel Kv1.1) (HUKI) (HBK1). Potassium voltage-gated channel subfamily A member 5 (Potassium channel Kv1.5) (HK2)	(HPCN1). Potassium voltage-gated channel subfamily H member 7 (Ether-a-go-go related gene potassium	UMAN channel 3) (HERG-3) (Ether-a-go-go related protein 3) (Eag related protein 3). JMAN Protein transport protein Sec24C (SEC24-related protein C). 6007826 rab escort protein-2 [Mus musculus] Renal sodium-dependent phosphate transport protein 2 (Sodium/phosphate cotransporter 2)	(Na(+)/Pi cotransporter 2) (Renal sodium-phosphate transport protein 2) (Renal Na(+)-dependent phosphate cotransporter 2).	Retinoic acid receptor alpha (RAR-alpha). Retinoic acid receptor gamma-1 (RAR-gamma-1).		Stalle acid officing 19-like fectin 3 preculsor (Signey-5) (Obesity Stricting process 2) (Ob-BP2) (CD33 antigen-like 2) (CD170 antigen). Sodium- and chloride-dependent glycine transporter 1 (GlyT-1) (GlyT-1).	(Sodium/faurocholate cotransporting polypeptide).
79 P11D_HUMAN 80 PIGO_HUMAN 81 940231	82 KPRA_HUMAN 83 PIT1_HUMAN 84 PKHD_HUMAN 85 PKHD_HUMAN 86 PKHD_HUMAN 87 P1L1_HUMAN	88 PKDR_HUMAN 89 PKD1_HUMAN	90 PAGT_HUMAN 91 DP1_HUMAN	92 CIK1_HUMAN	93 CIK5_HUMAN	94 KCH7_HUMAN 95 S24C_HUMAN 96 600782	97 NPT2 HUMAN	98 RRA_HUMAN 99 RRG1 HUMAN	100 RRG1_HUMAN 101 12053793	102 SIL5_HUMAN 103 S6A9_HUMAN	104 NTCP_HUMAN

835 PRSS <u>FAFP</u> PSLAKAG 525 EYGS <u>YRFP</u> PWAELLG ic	279 SLLM <u>FGFP</u> QSLPPHS 296 FDEL <u>FQFP</u> VVYDQLS 163 FSVD <u>YNFP</u> KKALVVT		570 YTVR <u>ETEP</u> EDPPLSP 570 YTVR <u>ETEP</u> DPPPLSP	767 GFGS <u>FRFP</u> SGNQGGA 498 FTMD <u>FCFP</u> FSPLQAF 1518 YILD <u>FQYP</u> FSAVQAF	351 HSKT <u>YHFP</u> KTVTQSL	228 ETCS <u>FCFP</u> ECRAPTQ 114 YRIR <u>FYFP</u> RWYCSGS 3)	184 NGGG <u>FLFP</u> PSYVPVV	186 CWAD <u>FGFP</u> WGPRAYL 1971 PYGV <u>FIFP</u> NKTRPLS 577 KSLC <u>FQYP</u> PVYVGKI 413 FFVI <u>FSFP</u> IASKDCI	1-2) 342 CGKG <u>FDFP</u> GSARIHE	Amino	Acid Target Sequence beta	213 TVGA <u>FYFP</u> TLLLIAL 42 DGGR <u>FKFP</u> DGVQNWP
Sodium/hydrogen exchanger 5 (Na(+)/H(+) exchanger 5) (NHE-5). Sodium-dependent proline transporter. Solute carrier family 21 member 11 (Sodium-independent organic anion transporter D) (Organic anion transporting polymentide D) (Organic anion transporter polymentide palated protein	 (OATP-RP3) (OATPRP3) (PGE1 transporter). Synaptotagmin X (SytX). Synaptotagmin XI (SytXI). Thyroid hormone receptor-associated protein complex 240 kDa component (Tran240) (Activator- 	recruited cofactor 250 kDa component) (ARC250). Thyroid hormone receptor-associated protein complex 240 kDa component (Trap240) (Activator-recruited cofactor 250 kDa component) (ABC250).			(Osteoclastogenesis inhibitory factor). Tumor necrosis factor receptor superfamily member 13B (Transmembrane activator and CAML.	interactor). Tyrosine-protein kinase JAK2 (EC 2.7.1.112) (Janus kinase 2) (JAK-2). UDP-glucuronosyltransferase 2B15 precursor, microsomal (EC 2.4.1.17) (UDPGT) (UDPGTH-3)			Zilic ilitger protein 44 (zilic ilitger protein NOA7) (Gonadoli opin ilitaddide u anscription repressor-z).		i 5-hydroxytryptamine 1B receptor (5-HT-1B) (Serotonin receptor) (5-HT-1D-beta) (Serotonin 1D beta	receptor) (S12). 5-hydroxytryptamine 2C receptor (5-HT-2C) (Serotonin receptor) (5HT-1C).
105 NAH5_HUMAN 106 S6A7_HUMAN	107 S21B_HUMAN 108 SYTA_MOUSE 109 SYTB_HUMAN	110 T240_HUMAN	112 TRIB_HUMAN 113 15277232	114 TERA_HUMAN 115 TUL2_HUMAN 116 TUSP_HUMAN	117 T11B_HUMAN	118 T13X_HUMAN 119 JAK2_HUMAN	120 UDBF_HUMAN	121 V1BR_HUMAN 122 16904210 123 WRN_HUMAN 124 WFS1_HUMAN	125 ZN44_HUMAN		Accession code	1 5H1B_HUMAN 2 5H2C_HUMAN

158 SLDIYNEPFDVQNCS 1057 GGEREPYPSFHWDPI 649 CAHWEQYPFDPSFDD 318 GVRSEGEPRPQQAAY 341 ELQQEGEPAPQTGDP 241 SYAKELYPTNALVTH 2649 TVFAENFPSLDALNT 329 TGYSEDFPFLEDSVK 61 VGTI FREPGGVSGEE	309 GAKA <u>FYYP</u> KEAGVPF 80 TLQI <u>FPFP</u> SSFSTL 598 FIQP <u>EFFP</u> RFSIGQR 89 TEIP <u>FEFP</u> LHLKGNK 1867 FNWR <u>FIFP</u> FDYLPAE 596 LYFH <u>FKFP</u> GTKTYID 687 SLFD <u>FIFP</u> GKLGTLP 48 EKFY <u>FAFP</u> GEILMRM	639 STDEFTWPKPNITSS 193 LEDFEVYPAEGPQIG 388 KKFKYLFPKFTLCWT 212 SFSSESYPENEMIYK 609 KWSRELFPLYAKEIH 81 GESEFLFPLYAKEIH 350 NYTEFKFPQIKAHPW 945 DASSFFFPPISSCPP 340 CFRDFCFPLKMRMER 46 NTRDEMFPGPNQMSG 3 GLLLFFFPAIFLEVS 9 RYPDFSFPYFPQDYF 751 RGAAFGFPGASPRAS
5-hydroxytryptamine 3 receptor precursor (5-HT-3) (Serotonin-gated ion channel receptor) (5-HT3R). Autism susceptibility gene 2 protein. Baculoviral IAP repeat-containing protein 1 (Neuronal apoptosis inhibitory protein). Brain-specific angiogenesis inhibitor 1 precursor. Brain-specific angiogenesis inhibitor 1 precursor. Cdk5 and abl enzyme substrate 2 (Interactor with cdk3 2) (Ik3-2). Cdk5 and abl enzyme substrate 2 (Interactor with cdk3 2) (Ik3-2). Noiliary dynein heavy chain 11 (Axonemal beta dynein heavy chain 11). Dimethylaniline monooxygenase [N-oxide forming] 5 (EC 1.14.13.8) (Hepatic flavin-containing monooxygenase 5) (FMO 5) (Dimethylaniline oxidase 5).		
3 5HT3_HUMAN 4 AUT2_HUMAN 5 BIR1_HUMAN 6 BRA1_HUMAN 7 BAI1_HUMAN 8 CBL2_HUMAN 9 DYHB_HUMAN 10 FMO5_HUMAN	12 DOPO_MOUSE 13 DSCA_HUMAN 14 DSCA_HUMAN 15 DSR3_HUMAN 16 DYSF_HUMAN 17 EPA7_HUMAN 18 ERC6_HUMAN 19 EAA3_HUMAN	21 FMR2_HUMAN 22 FMR2_HUMAN 23 FCMD_HUMAN 24 GAE_HUMAN 25 GAAT_HUMAN 26 SGCG_HUMAN 27 KG3A_HUMAN 27 KG3A_HUMAN 31 MGR1_HUMAN 31 MGR1_HUMAN 31 MGR1_HUMAN 33 MY15_HUMAN

431 FSPR <u>FPFP</u> TVPPAPG 413 FFVI <u>FSFP</u> IASKDCI	-	Amino Target Sequence Acid	1023 SLNP <u>FRFP</u> KEAASLF	922 VLPN <u>YLFP</u> TPSSYPY	907 MLPSYSFPSGTPNLP	843 PYPAFPFPYLDTFMT		661 EVCQFSYPQTPASPQ	
Williams-Beuren syndrome chromosome region 14 protein (WS basic-helix- loop-helix leucine zipper protein) (WS-bHLH) (MIx interactor). Wolframin.	TABLE 7: MISCELLANEOUS DEF DOMAIN-CONTAINING PROTEINS	Target Description	15425674 Per1 interacting protein of the suprachiamatic nucleus [Rattus norvegicus]	Period circadian protein 1 (Circadian pacemaker protein Rigui) (hPER).		Period circadian protein 3 (hPER3).	A-kinase anchor protein 11 (Protein kinase A anchoring protein 11) (PRKA11) (A kinase anchor	protein 220 kDa) (AKAP 220) (hAKAP220).	A-kinase anchor protein 3 (Protein kinase A anchoring protein 3) (PRKA3) (A-kinase anchor protein
62 WS14_HUMAN 63 WFS1_HUMAN		Accession Code	1 15425674	2 PER1 HUMAN	3 PER2_HUMAN	4 PER3 HUMAN	ı	5 AK11 HUMAN	I